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=> s promoter conversion

L1 6 PROMOTER CONVERSION

=> d 1-6 bib ab

L1 ANSWER 1 OF 6 MEDLINE  
AN 1998088888 MEDLINE  
DN 98088888  
TI Inducible expression of p21WAF-1/CIP-1/SDI-1 from a **promoter  
conversion** retroviral vector.  
AU Mrochen S; Klein D; Nikol S; Smith J R; Salmons B; Gunzburg W H  
CS Institute of Molecular Virology, GSF-National Research Center for

Environment and Health, Universitat Munchen, Germany.

SO JOURNAL OF MOLECULAR MEDICINE, (1997 Nov-Dec) 75 (11-12) 820-8.  
Journal code: B8C. ISSN: 0946-2716.

CY GERMANY: Germany, Federal Republic of

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 199804

EW 19980402

AB Constitutive, high-level expression of the potentially therapeutic WAF-1/CIP-1/SDI-1 gene is incompatible with cell growth. A **promoter conversion** retroviral vector carrying the WAF-1/CIP-1/SDI-1 gene under the transcriptional control of the glucocorticoid inducible promoter of mouse mammary tumor virus was used to infect human bladder carcinoma or feline kidney cells. Reduced cell growth due to a greater proportion of cells being in the G0/G1 phase of the cell cycle was observed when WAF-1/CIP-1/SDI-1 expression was activated by addition of glucocorticoid hormone. This system demonstrates the potential long-term therapeutic use of WAF-1/CIP-1/SDI-1 delivered by retroviral vectors for inhibiting the growth of rapidly proliferating cells. Moreover, the conditional expression of genes such as WAF-1/CIP-1/SDI-1 from such retroviral vectors may facilitate analysis of their function.

L1 ANSWER 2 OF 6 MEDLINE

AN 96228295 MEDLINE

DN 96228295

TI Importance of low affinity Elf-1 sites in the regulation of lymphoid-specific inducible gene expression.

AU John S; Marais R; Child R; Light Y; Leonard W J

CS Laboratory of Molecular Immunology, National Heart, Lung, and Blood Institute, National Institutes of Health, Bethesda, Maryland 20892, USA.

SO JOURNAL OF EXPERIMENTAL MEDICINE, (1996 Mar 1) 183 (3) 743-50.  
Journal code: I2V. ISSN: 0022-1007.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals; Cancer Journals

EM 199609

AB Elf-1 is an Ets family transcription factor that regulates a number of inducible lymphoid-specific genes, including those encoding interleukin 3 (IL-3), granulocyte/macrophage colony-stimulating factor (GM-CSF), and the IL-2 receptor (IL-2R) alpha chain. A minimal oligonucleotide spanning the IL-2R alpha Elf-1 site (-97/-84) bound Elf-1 poorly, but binding activity markedly increased when this oligonucleotide was multimerized or flanking sequences were added. This result is consistent with the requirement of accessory proteins for efficient Elf-1 binding, as has been demonstrated for the GM-CSF and IL-3 promoters. A binding site selection analysis revealed the optimal Elf-1 consensus motif to be A(A/t)(C/a)CCGGAAGT(A/S), which is similar to the consensus motif for the related Drosophila E74 protein. This minimal high affinity site could bind Elf-1 and functioned as a stronger transcription element than the -97/-84 IL-2R alpha oligonucleotide when cloned upstream of a heterologous promoter. In contrast, in the context of the IL-2R alpha **promoter, conversion** of the naturally occurring low affinity Elf-1 site to an optimal site decreased inducible activation of a reporter construct in Jurkat

cells. This finding may be explained by the observation that another Ets family protein, ER GB/Fli-1, can efficiently bind only to the optimal site, and in this context, interferes with Elf-1 binding. Therefore, high affinity Elf-1 sites may lack sufficient binding specificity, whereas naturally occurring low affinity sites presumably favor the association of Elf-1 in the context of accessory proteins. These findings offer an explanation for the lack of optimal sites in any of the known Elf-1-regulated genes.

L1 ANSWER 3 OF 6 MEDLINE  
 AN 93113085 MEDLINE  
 DN 93113085  
 TI Conversion of dethiobiotin to biotin in cell-free extracts of *Escherichia coli*.  
 AU Ifuku O; Kishimoto J; Haze S; Yanagi M; Fukushima S  
 CS Shiseido Research Center, Yokohama, Japan..  
 SO BIOSCIENCE, BIOTECHNOLOGY, AND BIOCHEMISTRY, (1992 Nov) 56 (11) 1780-5.  
 Journal code: BDP. ISSN: 0916-8451.  
 CY Japan  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS B  
 EM 199304  
 AB We constructed the plasmid pTTB151 in which the *E. coli* bioB gene was expressed under the control of the tac **promoter**. **Conversion** of dethiobiotin to biotin was demonstrated in cell-free extracts of *E. coli* carrying this plasmid. The requirements for this biotin-forming reaction included fructose-1,6-bisphosphate, Fe<sup>2+</sup>, S-adenosyl-L-methionine, NADPH, and KCl, as well as dethiobiotin as the substrate. The enzymes were partially purified from cell-free extracts by a procedure involving ammonium sulfate fractionation. Our results suggest that an unidentified enzyme(s) besides the bioB gene product is obligatory for the conversion of dethiobiotin to biotin.

L1 ANSWER 4 OF 6 BIOSIS COPYRIGHT 1998 BIOSIS  
 AN 98:76483 BIOSIS  
 DN 01076483  
 TI Inducible expression of p21-WAF-1-CIP-1-SDI-1 from a **promoter conversion** retroviral vector.  
 AU Mrochen S; Klein D; Nikol S; Smith J R; Salmons B; Guenzburg W H  
 CS Inst. Virol., Univ. Veterinary Sci., Josef-Baumann-Gasse 1, A-1210 Vienna, Austria  
 SO Journal of Molecular Medicine (Berlin) 75 (11-12). 1997. 820-828. ISSN: 0946-2716  
 LA English  
 AB Constitutive, high-level expression of the potentially therapeutic WAF-1/CIP-1/SDI-1 gene is incompatible with cell growth. A **promoter conversion** retroviral vector carrying the WAF-1/CIP-1/SDI-1 gene under the transcriptional control of the glucocorticoid inducible promoter of mouse mammary tumor virus was used to infect human bladder carcinoma or feline kidney cells. Reduced cell growth due to a greater proportion of cells being in the G-0/G-1 phase of the cell cycle was observed when WAF-1/CIP-1/SDI-1-expression was activated by addition of glucocorticoid hormone. This system demonstrates the potential long-term therapeutic use of WAF-1/CIP-1/SDI-1 delivered by retroviral vectors for inhibiting the growth of rapidly proliferating cells. Moreover, the conditional expression of genes such as

WAF-1/CIP-1/SDI-1 from such retroviral vectors may facilitate analysis of their function.

L1 ANSWER 5 OF 6 BIOSIS COPYRIGHT 1998 BIOSIS  
AN 96:238164 BIOSIS  
DN 98786293  
TI Importance of low affinity Elf-1 sites in the regulation of lymphoid-specific inducible gene expression.  
AU John S; Marais R; Child R; Light Y; Leonard W J  
CS Lab. Molecular Immunology, Natl. Heart, Lung, and Blood Inst., Natl. Inst. Health, Bethesda, MD 20892, USA  
SO Journal of Experimental Medicine 183 (3). 1996. 743-750. ISSN: 0022-1007  
LA English  
AB Elf-1 is an Ets family transcription factor that regulates a number of inducible lymphoid-specific genes, including those encoding interleukin 3 (IL-3), granulocyte/macrophage colony-stimulating factor (GM-CSF), and the IL-2 receptor (IL-2R) alpha chain. A minimal oligonucleotide spanning the IL-2-alpha Elf-1 site (-97/-84) bound Elf-1 poorly, but binding activity markedly increased when this oligonucleotide was multimerized or flanking sequences were added. This result is consistent with the requirement of accessory proteins for efficient Elf-1 binding, as has been demonstrated for the GM-CSF and IL-3 promoters. A binding site selection analysis revealed the optimal Elf-1 consensus motif to be A(A/t)(C/a)CCGGAAGT(A/S), which is similar to the consensus motif for the related Drosophila E74 protein. This minimal high affinity site could bind Elf-1 and functioned as a stronger transcription element than the -97/-84 IL2R-alpha oligonucleotide when cloned upstream of a heterologous promoter. In contrast, in the context of the IL-2R-alpha promoter, conversion of the naturally occurring low affinity Elf-1 site to an optimal site decreased inducible activation of a reporter construct in Jurkat cells. This finding may be explained by the observation that another Ets family protein, ERGB/Fli-1, can efficiently bind only to the optimal site, and in this context, interferes with Elf-1 binding. Therefore, high affinity Elf-1 sites may lack sufficient binding specificity, whereas naturally occurring low affinity sites presumably favor the association of Elf-1 in the context of accessory proteins. These findings offer an explanation for the lack of optimal sites in any of the known Elf-1-regulated genes.

L1 ANSWER 6 OF 6 BIOSIS COPYRIGHT 1998 BIOSIS  
AN 93:168763 BIOSIS  
DN BA95:89813  
TI CONVERSION OF DETHIOBIOTIN TO BIOTIN IN CELL-FREE EXTRACTS OF ESCHERICHIA-COLI.  
AU IFUKU O; KISHIMOTO J; HAZE S-I; YANAGI M; FUKUSHIMA S  
CS SHISEIDO RES. CENT., 1050, NIPPA-CHO, KOHOKU-KU, YOKOHAMA 223, JAPAN.  
SO BIOSCI BIOTECHNOL BIOCHEM 56 (11). 1992. 1780-1785. CODEN: BBBIEJ  
LA English  
AB We constructed the plasmid pTTB151 in which are E. coli bioB gene was expressed under the control of the tac promoter.  
Conversion of dethiobiotin to biotin was demonstrated in cell-free extracts of E. coli carrying this plasmid. The requirements for this biotin-forming reaction included fructose-1,6-bisphosphate, Fe2+, S-adenosyl-L-methione, NADPH, and KCl, as well as dethiobiotin as the substrate. The enzymes were partially purified from cell-free extracts by a procedure involving ammonium sulfate fractionation. Our results suggest that an unidentified enzyme(s) besides the bioB gene

product is obligatory for the conversion of dethiobiotin in biotin.

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=> s self inactivating and retrovir?

L1 43 SELF INACTIVATING AND RETROVIR?

=> s l1 range=,1994

L2 24 L1

=> d 1-10 bib ab

L2 ANSWER 1 OF 24 MEDLINE  
AN 93211392 MEDLINE  
DN 93211392

TI Stable expression of antibiotic resistance genes using a promoter fragment of the U1 snRNA gene.  
 AU Asselbergs F A; Pronk R  
 CS Biotechnology Department, CIBA-GEIGY Ltd., Basle, Switzerland..  
 SO MOLECULAR BIOLOGY REPORTS, (1993 Feb) 17 (2) 101-14.  
 Journal code: NGW. ISSN: 0301-4851.  
 CY Netherlands  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199307  
 AB As U1 snRNA is produced in all mammalian cell types, antibiotic resistance genes driven by this promoter would be ideally suited as genetic selection markers. However, although the U1 snRNA gene is transcribed by RNA polymerase II, its native product is not a messenger RNA, but a splicing cofactor. To test whether this promoter could nevertheless produce a functional mRNA, sensitive reporter genes expressing resistance to the antibiotics hygromycin-B and bleomycin were constructed with either the U1 snRNA promoter or the SV40 early promoter. Resistant cell lines could only be obtained with constructs equipped with a functional polyadenylation signal. With the U1 snRNA promoter about three times fewer colonies were obtained than with the SV40 early promoter. Another potential advantage of the U1 snRNA promoter is that, in contrast to the promoters commonly used to express genetic selection markers, the enhancer-like element contained in the U1 snRNA promoter had only a minimal stimulative effect, only detectable with the most sensitive methods, on an adjacent mRNA-producing gene. The U1 snRNA promoter was also capable of expressing bleomycin resistance in the context of a **self-inactivating retrovirus** vector, whereby it was discovered that the mouse 3T3 cells used in this experiment were 10 times more sensitive to bleomycin than human or hamster cell lines.

L2 ANSWER 2 OF 24 MEDLINE  
 AN 93139782 MEDLINE  
 DN 93139782  
 TI Importance of 3' non-coding sequences for efficient **retrovirus**-mediated gene transfer in avian cells revealed by **self-inactivating** vectors.  
 AU Flamant F; Aubert D; Legrand C; Cosset F L; Samarut J  
 CS Laboratoire de Biologie Moleculaire et Cellulaire, INRA-CNRS UMR 49, France.  
 SO JOURNAL OF GENERAL VIROLOGY, (1993 Jan) 74 ( Pt 1) 39-46.  
 Journal code: I9B. ISSN: 0022-1317.  
 CY ENGLAND: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199304  
 AB Avian leukosis virus-derived vectors were constructed with an internal transcriptional promoter and various 3' non-coding sequences. Deletions were introduced into the downstream U3 long terminal repeat (LTR) to obtain self-inactivation of LTR-mediated transcription after one round of replication. However, 3' non-coding sequences appeared to determine not only self-inactivation of the vectors but also gene transfer efficiency. Further analysis revealed the influence of these sequences on both internal gene expression and RNA packaging. One construct permitted gene transfer while inactivating 5' LTR-promoted transcription.

L2 ANSWER 3 OF 24 MEDLINE  
 AN 93114447 MEDLINE  
 DN 93114447  
 TI An improved **retroviral** vector for assaying promoter activity. Analysis of promoter interference in pIP211 vector.  
 AU Nakajima K; Ikenaka K; Nakahira K; Morita N; Mikoshiba K  
 CS Institute for Protein Research, Osaka University, Japan..  
 SO FEBS LETTERS, (1993 Jan 4) 315 (2) 129-33.  
 Journal code: EUH. ISSN: 0014-5793.  
 CY Netherlands  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199304  
 AB We recently developed a novel promoter assay system using a **retroviral** vector (pIP200 series). Transcription from the internal promoter, which had been inserted for the promoter assay, was shown to be interfered with by transcription from the upstream long terminal repeat (LTR). Here we report a new high-titer '**self-inactivating**' vector, in which transcription interference was virtually eliminated. This new vector was constructed by introducing only a very minor mutation into the 'TATA box' in the 3'-LTR. This mutation was successfully transferred to the 5'-LTR after reverse transcription, yielding a provirus incapable of transcribing viral RNA. The viral titer was not reduced by the mutation, permitting general application of this virus.

L2 ANSWER 4 OF 24 MEDLINE  
 AN 91255559 MEDLINE  
 DN 91255559  
 TI MoMuLV-derived **self-inactivating** **retroviral** vectors possessing multiple cloning sites and expressing the resistance to either G418 or hygromycin B.  
 AU Marty L; Roux P; Royer M; Piechaczyk M  
 CS Laboratoire de Biologie Moleculaire, URA CNRS 1191 Genetique Moleculaire, Montpellier, France..  
 SO BIOCHIMIE, (1990 Dec) 72 (12) 885-7.  
 Journal code: A14. ISSN: 0300-9084.  
 CY France  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199109  
 AB To facilitate cloning procedures in recombinant murine leukemia virus-derived **retroviruses**, we have constructed vectors that both carry a polylinker with multiple restriction sites and express resistance to either G418 or hygromycin B. Our vectors are **self-inactivating retroviruses** that suppress interferences between LTR enhancers and internal promoters and avoid transcriptional stimulation of host cell genes. They can also be used as expression vectors in direct transfection assays, since no translation initiation codon lies between the 5' LTR and the cloning polylinker.

L2 ANSWER 5 OF 24 MEDLINE  
 AN 91202569 MEDLINE  
 DN 91202569  
 TI Promoter interactions in **retrovirus** vectors introduced into fibroblasts and embryonic stem cells.



AU Soriano P; Friedrich G; Lawinger P  
 CS Howard Hughes Medical Institute, Baylor College of Medicine,  
 Houston, Texas 77030.  
 NC RR05425 (NCRR)  
 R01-HD24875 (NICHD)  
 SO JOURNAL OF VIROLOGY, (1991 May) 65 (5) 2314-9.  
 Journal code: KCV. ISSN: 0022-538X.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199107  
 AB The activity of the Moloney murine leukemia virus promoter is  
 restricted in mouse embryonic stem cells. Gene expression with  
**retrovirus** vectors can be achieved in these cells if  
 internal promoters are used. To address the possible influence of  
 the viral enhancer sequences on expression from the internal  
 promoter, we have constructed high-titer, **self-**  
**inactivating retrovirus** vectors which delete viral  
 regulatory sequences upon integration in the host genome. We show  
 that deleting most of the viral enhancer sequences has no  
 significant effect on viral titer. This enhancer deletion leads to  
 either an increase or a decrease in the amount of RNA transcribed  
 from the internal promoter, but no consistent change can be found  
 with any type of vector. The same changes in expression from the  
 internal promoter observed in embryonic stem cells are also observed  
 in 3T3 fibroblast cells, in which the viral promoter is active.  
 These results indicate that viral regulatory elements influence  
 expression from an internal promoter independently of expression  
 from the virus promoter.

L2 ANSWER 6 OF 24 MEDLINE  
 AN 91201400 MEDLINE  
 DN 91201400  
 TI Inhibition of proliferation of primary avian fibroblasts through  
 expression of histone H5 depends on the degree of phosphorylation of  
 the protein.

AU Aubert D; Garcia M; Benchaibi M; Poncet D; Chebloune Y; Verdier G;  
 Nigon V; Samarut J; Mura C V  
 CS Laboratoire de Biologie Moléculaire et Cellulaire, UMR 13 Centre  
 National de la Recherche Scientifique, Lyon, France.  
 SO JOURNAL OF CELL BIOLOGY, (1991 May) 113 (3) 497-506.  
 Journal code: HMV. ISSN: 0021-9525.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199107  
 AB To obtain stable and constitutive expression of histone H5 at levels  
 comparable to those observed in normal chicken erythrocytes, an  
 avian **self-inactivating retroviral**  
 vector was used to transfer the H5 gene into cells which do not  
 express this protein. The vector, pDAH5, was obtained by removing  
 the CAAT and TATA boxes of the 3'LTR of the avian leukosis virus  
 RAV-2 and inserting the H5 sequence. Infection of QT6 quail cells  
 with the recombinant virus (DAH5) led to the stable integration of  
 the foreign H5 gene at low copy number, to the formation of  
 correctly initiated mRNA transcripts and to the production of H5  
 protein. The amount of H5 expressed was equivalent to that of a  
 mature chicken erythrocyte. Expression of histone H5 in DAH5

transformed cells, such as QT6 or AEV-ES4, transformed chicken embryo fibroblasts had only slight effects on the growth rate and did not inhibit cell replication. Conversely, the effect of H5 expression on normal quail and chicken fibroblasts was dramatic: cells acquired the aspect of quiescent fibroblasts, grew very slowly, and nuclei looked compacted, often extruded from the cell. The H5 histone produced in QT6-transformed cells was found to be phosphorylated while in normal chicken fibroblasts the protein lacked this posttranslational modification. It is proposed that the chromatin-condensing role of histone H5 is inhibited by its phosphorylation.

L2 ANSWER 7 OF 24 MEDLINE  
AN 90355975 MEDLINE  
DN 90355975  
TI Functional analysis and nucleotide sequence of the promoter region of the murine hck gene.  
AU Lock P; Stanley E; Holtzman D A; Dunn A R  
CS Ludwig Institute for Cancer Research, Melbourne Tumour Biology Branch, Parkville, Victoria, Australia.  
SO MOLECULAR AND CELLULAR BIOLOGY, (1990 Sep) 10 (9) 4603-11.  
Journal code: NGY. ISSN: 0270-7306.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals  
OS GENBANK-M34946  
EM 199011  
AB The structure and function of the promoter region and exon 1 of the murine hck gene have been characterized in detail. RNase protection analysis has established that hck transcripts initiate from heterogeneous start sites located within the hck gene. Fusion gene constructs containing hck 5'-flanking sequences and the bacterial Neor gene have been introduced into the hematopoietic cell lines FDC-P1 and WEHI-265 by using a **self-inactivating retroviral** vector. The transcriptional start sites of the fusion gene are essentially identical to those of the endogenous hck gene. Analysis of infected WEHI-265 cell lines treated with bacterial lipopolysaccharide (LPS) reveals a 3- to 5-fold elevation in the levels of endogenous hck mRNA and a 1.4- to 2.6-fold increase in the level of Neor fusion gene transcripts, indicating that hck 5'-flanking sequences are capable of conferring LPS responsiveness on the Neor gene. The 5'-flanking region of the hck gene contains sequences similar to an element which is thought to be involved in the LPS responsiveness of the class II major histocompatibility gene A alpha k. A subset of these sequences are also found in the 5'-flanking regions of other LPS-responsive genes. Moreover, this motif is related to the consensus binding sequence of NF-kappa B, a transcription factor which is known to be regulated by LPS.

L2 ANSWER 8 OF 24 MEDLINE  
AN 90269622 MEDLINE  
DN 90269622  
TI Construction and hormone regulation of a novel **retroviral** vector.  
AU Mee P J; Brown R  
CS CRC Department of Medical Oncology, Glasgow University, U.K.  
SO GENE, (1990 Apr 16) 88 (2) 289-92.  
Journal code: FOP. ISSN: 0378-1119.  
CY Netherlands

DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199009  
 AB We report the analysis of a **self-inactivating retroviral** vector, constructed to allow inducible gene expression of inserted sequences from the mouse mammary tumour virus hormonal response element. The cloning strategy has been designed to allow for ease of insertion of the genes of interest. The vector contains the aph gene, allowing geneticin-resistance selection in mammalian cells. We have characterised dexamethasone (Dex)-induced increase in gene expression using the reporter gene encoding chloramphenicol acetyltransferase (CAT) inserted into the **retroviral** vector. We observe low basal levels of CAT activity in infected cells which is increased up to 50-fold by induction with Dex. The induction of pooled clones is 13.3-fold. Variation in Dex-induced CAT activity is observed in independently infected clones, which is not explained by proviral copy number.

L2 ANSWER 9 OF 24 MEDLINE

AN 90014804 MEDLINE

DN 90014804

TI Germ line c-myc is not down-regulated by loss or exclusion of activating factors in myc-induced macrophage tumors.

AU Mango S E; Schuler G D; Steele M E; Cole M D

CS Lewis Thomas Laboratory, Department of Biology, Princeton University, New Jersey 08544-1014..

SO MOLECULAR AND CELLULAR BIOLOGY, (1989 Aug) 9 (8) 3482-90.

Journal code: NGY. ISSN: 0270-7306.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

OS GENBANK-M28065; GENBANK-M28066

EM 199001

AB As in tumors with c-myc chromosomal translocations, c-myc **retrovirus**-induced monocyte tumors constitutively express an activated form of c-myc (the proviral gene), whereas the normal endogenous c-myc genes are transcriptionally silent. Treatment of these **retrovirus**-induced tumor cells with a number of bioactive chemicals and growth factors that are known to induce c-myc expression in cells of the monocyte lineage failed to induce the endogenous c-myc gene. In contrast, the same treatments induced the c-fos gene in both tumors and a control macrophage line. To investigate c-myc suppression further, a normal copy of the human c-myc gene was introduced into tumor and control cell lines by using a **retrovirus** with **self-inactivating** long terminal repeats. This transduced normal gene was expressed at equivalent levels in all cells, regardless of the state of endogenous c-myc gene expression, and was strongly induced by agents that induce the normal gene in the control cells. These results indicate that the signal transduction pathways that normally activate the c-myc gene are functional in myc-induced tumor cells and suggest that endogenous c-myc is actively suppressed. An examination of the c-myc locus itself showed that the lack of transcriptional activity correlated with the absence of several prominent DNase I-hypersensitive sites in the 5'-flanking region of the gene but without loss of general DNase sensitivity. Furthermore, analysis of 22 methylation-sensitive restriction enzyme sites in the 5'-flanking region, first exon, and first intron indicated that the

silent c-myc genes remained in the same unmethylated state as did actively expressed genes. Thus, c-myc suppression does not appear to result from the most frequently described mechanisms of gene inactivation.

L2 ANSWER 10 OF 24 MEDLINE  
AN 89315828 MEDLINE  
DN 89315828  
TI Analysis of mammalian cell genetic regulation in situ by using **retrovirus**-derived "portable exons" carrying the Escherichia coli lacZ gene.  
AU Brenner D G; Lin-Chao S; Cohen S N  
CS Department of Genetics, Stanford University School of Medicine, CA 94305-5120.  
NC GM 27241 (NIGMS)  
SO PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, (1989 Jul) 86 (14) 5517-21.  
Journal code: PV3. ISSN: 0027-8424.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 198910  
AB **Self-inactivating** derivatives of Moloney murine leukemia **retrovirus** containing the Escherichia coli lacZ gene were used to detect and study the regulation of transcription initiated at chromosomally located promoters in mouse fibroblasts. The introduction of splice acceptor sites in all three translational reading frames relative to lacZ and the inclusion of an in-frame ATG translation start codon in one construct allowed synthesis of beta-galactosidase fusion proteins upon insertion of **retrovirus** vectors containing lacZ into introns 3' to either protein-coding or noncoding exons. Selection of lacZ-expressing cells by fluorescence-activated cell sorting and the analysis of beta-galactosidase production after serum deprivation has yielded lines in which lacZ was fused to genes induced by growth arrest in the G0 state.

=> d 11-24 bib ab

L2 ANSWER 11 OF 24 MEDLINE  
AN 86205863 MEDLINE  
DN 86205863  
TI **Self-inactivating retroviral** vectors designed for transfer of whole genes into mammalian cells.  
AU Yu S F; von Ruden T; Kantoff P W; Garber C; Seiberg M; Ruther U; Anderson W F; Wagner E F; Gilboa E  
SO PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, (1986 May) 83 (10) 3194-8.  
Journal code: PV3. ISSN: 0027-8424.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 198608  
AB A **retrovirus**-derived vector called **self-inactivating** (SIN) vector was designed for the transduction of whole genes into mammalian cells. SIN vectors contain a deletion of 299 base pairs in the 3' long terminal repeat (LTR), which

includes sequences encoding the enhancer and promoter functions. When viruses derived from such vectors were used to infect NIH 3T3 cells, the deletion was transferred to the 5' LTR, resulting in the transcriptional inactivation of the provirus in the infected cell. Introduction of a hybrid gene (human metallothionein-promoted c-fos) into cells via a SIN vector was not associated with rearrangements and led to the formation of an authentic mRNA transcript, which in some cases was induced by cadmium. SIN vectors should be particularly useful in gene transfer experiments designed to study the regulated expression of genes in mammalian cells. Absence of enhancer and promoter sequences in both LTRs of the integrated provirus should also minimize the possibility of activating cellular oncogenes and may provide a safer alternative to be used in human gene therapy.

L2 ANSWER 12 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 94:545441 BIOSIS

DN 98004989

TI Improved **self-inactivating retroviral** vectors derived from spleen necrosis virus.

AU Olson P; Nelson S; Dornburg R

CS Dep. Molecular Genetics Microbiol., Robert Wood Johnson Med. Sch., Univ. Medicine Dentistry New Jersey, 675 Hoes Lane, Piscataway, NJ 08854, USA

SO Journal of Virology 68 (11). 1994. 7060-7066. ISSN: 0022-538X

LA English

AB **Self-inactivating (SIN) retroviral**

vectors contain a deletion spanning most of the right long terminal repeat's (LTR's) U3 region. Reverse transcription copies this deletion to both LTRs. As a result, there is no transcription from the 5' LTR, preventing further replication. Many previously developed SIN vectors, however, had reduced titers or were genetically unstable. Earlier, we reported that certain SIN vectors derived from spleen necrosis virus (SNV) experienced reconstitution of the U3-deleted LTR at high frequencies. This reconstitution occurred on the DNA level and appeared to be dependent on defined vector sequences. To study this phenomenon in more detail, we developed an almost completely U3-free **retroviral** vector. The promoter and enhancer of the left LTR were replaced with those of the cytomegalovirus immediate-early genes. This promoter swap did not impair the level of transcription or alter its start site. Our data indicate that SNV contains a strong initiator which resembles that of human immunodeficiency virus. We show that the vectors replicate with efficiencies similar to those of vectors possessing two wild-type LTRs. U3-deleted vectors carrying the hygromycin B phosphotransferase gene did not observably undergo LTR reconstitution, even when replicated in helper cells containing SNV-LTR sequences. However, vectors carrying the neomycin resistance gene did undergo LTR reconstitution with the use of homologous helper cell LTR sequences as template. This supports our earlier finding that sequences within the neomycin resistance gene can trigger recombination.

L2 ANSWER 13 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 93:251520 BIOSIS

DN BA95:130695

TI STABLE EXPRESSION OF ANTIBIOTIC RESISTANCE GENES USING A PROMOTER FRAGMENT OF THE U1 SNRNA GENE.

AU ASSELBERGS F A M; PRONK R

CS BIOTECHNOLOGY DEP., CIBA-GEIGY LTD., K681-442 CH-4002 BASLE, SWITZ.  
SO MOL BIOL REP 17 (2). 1993. 101-114. CODEN: MLBRBU ISSN: 0301-4851

LA English

AB As U1 snRNA is produced in all mammalian cell types, antibiotic resistance genes driven by this promoter would be ideally suited as genetic selection markers. However, although the U1 snRNA gene is transcribed by RNA polymerase II, its native product is not a messenger RNA, but a splicing cofactor. To test whether this promoter could nevertheless produce a functional mRNA, sensitive reporter genes expressing resistance to the antibiotics hygromycin-B and bleomycin was constructed with either the U1 snRNA promoter or the SV40 early promoter. Resistant cell lines could only be obtained with constructs equipped with a functional polyadenylation signal. With the U1 snRNA promoter about three times fewer colonies were obtained than with the SV40 promoter. Another potential advantage of the U1 snRNA promoter is that, in contrast to the promoter commonly used to express genetic selection markers, the enhancer-like element contained in the U1 snRNA promoter had only a minimal stimulative effect, only detectable with the most sensitive methods, on an adjacent mRNA-producing gene. The U1 snRNA promoter may also be capable of expressing bleomycin resistance in the context of a **self-inactivating retrovirus** vector, whereby it was discovered that the mouse 3T3 cells used in this experiment were 10 times more sensitive to bleomycin than human or hamster cell lines.

L2 ANSWER 14 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 93:164489 BIOSIS

DN BA95:85539

TI IMPORTANCE OF 3' NON-CODING SEQUENCES FOR EFFICIENT

**RETROVIRUS-MEDIATED GENE TRANSFER IN AVIAN CELLS REVEALED BY SELF-INACTIVATING VECTORS.**

AU FLAMANT F; AUBERT D; LEGRAND C; COSSET F-L; SAMARUT J

CS LAB. DE BIOL. MOL. CELLULAIRE, INRA-CNRS UMR 49, ECOLE NORMALE SUPERIEURE DE LYON, 46 ALLEE ITALIE, 69364 LYON CEDEX 07, FR.

SO J GEN VIROL 74 (1). 1993. 39-46. CODEN: JGVIAIY ISSN: 0022-1317

LA English

AB Avian leukosis virus-derived vectors were constructed with an internal transcriptional promoter and various 3' non-coding sequences. Deletions were introduced into the downstream U3 long terminal repeat (LTR) to obtain self-inactivation of LTR-mediated transcription after one round of replication. However, 3' non-coding sequences appeared to determine not only self-inactivation of the vectors but also gene transfer efficiency. Further analysis revealed the influence of these sequences on both internal gene expression and RNA packaging. One construct permitted gene transfer while inactivating 5' LTR-promoted transcription.

L2 ANSWER 15 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 93:140830 BIOSIS

DN BA95:73630

TI AN IMPROVED **RETROVIRAL** VECTOR FOR ASSAYING PROMOTER

ACTIVITY ANALYSIS OF PROMOTER INTERFERENCE IN PIP211 VECTOR.

AU NAKAJIMA K; IKENAKA K; NAKAHIRA K; MORITA N; MIKOSHIBA K

CS DIV. REGULATION MACROMOLECULAR FUNCTION, INST. PROTEIN RES., OSAKA UNIV., 3-2 YAMADAOKA, SUITA, OSAKA 565, JAPAN.

SO FEBS (FED EUR BIOCHEM SOC) LETT 315 (2). 1993. 129-133. CODEN: FEBLAL ISSN: 0014-5793

LA English

AB We recently developed a novel promoter assay system using a **retroviral** vector (pip200 series). Transcription from the internal promoter, which had been inserted for the promoter assay, was shown to be interfered with by transcription from the upstream

long terminal repeat (LTR). Here we report a new high-titer '**self-inactivating**' vector, in which transcription interference was virtually eliminated. This new vector was constructed by introducing only a very minor mutation into the 'TATA box' in the 3'-LTR. This mutation was successfully transferred to the 5'-LTR after reverse transcription, yielding a provirus incapable of transcribing viral RNA. The viral titer was not reduced by the mutation, permitting general application of this virus.

L2 ANSWER 16 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 92:434312 BIOSIS

DN BA94:86437

TI GENE-SEARCH VIRUSES AND FACS-GAL PERMIT THE DETECTION ISOLATION AND CHARACTERIZATION OF MAMMALIAN CELLS WITH IN-SITU FUSIONS BETWEEN CELLULAR GENES AND ESCHERICHIA-COLI LACZ.

AU KERR W G; HERZENBERG L A

CS DEP. GENETICS, STANFORD UNIV. SCH. MED., STANFORD, CALIF. 94305.

SO METHODS (ORLANDO) 2 (3). 1991. 261-271. CODEN: MTHDE9 ISSN: 1046-2023

LA English

AB We describe **self-inactivating**

**retroviruses** that will activate expression of the receptor gene, Escherichia coli lacZ, upon integration within a cellular gene (Gensrl) or near a cellular enhancer (Enhsrc1), referred to collectively as gene-search viruses. If the Gensrl virus integrates within an intron of a gene, splicing of a lacZ neo-exon to an upstream cellular exon can occur, resulting in transcriptional and translational fusion between E. coli lacZ and a cellular gene. The Enhsrc1 **retrovirus** generates a provirus that lacks the LTR enhancer region and thus is dependent upon flanking cellular enhancers to activate expression of lacZ. Fluorescence-activated cell sorting permits mammalian cells infected with the gene-search viruses that contain gene fusions between lacZ and cellular transcription control elements to be isolated as a population or as clones of single cells. Clones can be analyzed via a rapid, sensitive assay for .beta.-galactosidase activity carried out in 96-well plates, permitting clones with integrations in conditionally expressed genes to be identified. This approach has led to isolation of gene-search virus integrations in developmentally regulated genes and loci. In addition, lacZ+ clones derived from Gensrl infections can be screened via a histochemical stain (X-gal) for subcellular targeting of .beta.-gal activity. Molecular characterization of such Gensrl integrations could allow identification of mammalian proteins with specific subcellular localizations. Finally, we demonstrate the potential of gene-search viruses for obtaining expression of lacZ in normal cells both in vitro and in vivo.

L2 ANSWER 17 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 91:271386 BIOSIS

DN BA92:4001

TI PROMOTER INTERACTIONS IN **RETROVIRUS** VECTORS INTRODUCED INTO FIBROBLASTS AND EMBRYONIC STEM CELLS.

AU SORIANO P; FRIEDRICH G; LAWINGER P

CS HOWARD HUGHES MEDICAL INSTITUTE, BAYLOR COLLEGE MEDICINE, HOUSTON, TEX. 77030.

SO J VIROL 65 (5). 1991. 2314-2319. CODEN: JOVIAM ISSN: 0022-538X

LA English

AB The activity of the Moloney murine leukemia virus promoter is restricted in mouse embryonic stem cells. Gene expression with **retrovirus** vectors can be achieved in these cells if internal

promoters are used. To address the possible influence of the viral enhancer sequences on expression from the internal promoter, we have constructed high-titer, **self-inactivating retrovirus** vectors which delete viral regulatory sequences upon integration in the host genome. We show that deleting most of the viral enhancer sequences has no significant effect on viral titer. This enhancer deletion lead to either an increase or a decrease in the amount of RNA transcribed from the internal promoter, but no consistent change can be found with any type of vector. The same changes in expression from the internal promoter observed in embryonic stem cells are also observed in 3T3 fibroblast cells, in which the viral promoter is active. These results indicate that viral regulatory elements influence expression from an internal promoter independently of expression from the virus promoter.

L2 ANSWER 18 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS  
 AN 91:268135 BIOSIS  
 DN BA92:750  
 TI INHIBITION OF PROLIFERATION OF PRIMARY AVIAN FIBROBLASTS THROUGH EXPRESSION OF HISTONE H5 DEPENDS ON THE DEGREE OF PHOSPHORYLATION OF THE PROTEIN.  
 AU AUBERT D; GARCIA M; BENCHAIBI M; PONCET D; CHEBLOUNE Y; VERDIER G; NIGON V; SAMARUT J; MURA C V  
 CS LAB. DE BIOLOGIE MOLECULAIRE ET CELLULAIRE, UMR 13 CENTRE NATL. DE LA RECHERCHE SCIENTIFIQUE, ECOLE NORMALE SUPERIEURE DE LYON, 69364 LYON CEDEX 07, FRANCE.  
 SO J CELL BIOL 113 (3). 1991. 497-506. CODEN: JCLBA3 ISSN: 0021-9525  
 LA English  
 AB To obtain stable and constitutive expression of histone H5 at levels comparable to those observed in normal chicken erythrocytes, an avian **self-inactivating retroviral** vector was used to transfer the H5 gene into cells which do not express this protein. The vector, pDAH5, was obtained by removing the CAAT and TATA boxes of the 3LTR of the avian leukosis virus RAV-2 and inserting the H5 sequence. Infection of QT6 quail cells with the recombinant virus (DAH5) led to the stable integration of the foreign H5 gene at low copy number, to the formation of correctly initiated mRNA transcripts and to the production of H5 protein. The amount of H5 expressed was equivalent to that of a mature chicken erythrocyte. Expression of histone H5 in DAH5 transformed cells, such as QT6 or AEV-ES4, transformed chicken embryo fibroblasts had only slight effects on the growth rate and did not inhibit cell replication. Conversely, the effect of H5 expression on normal quail and chicken fibroblasts was dramatic: cells acquired the aspect of quiescent fibroblasts, grew very slowly, and nuclei looked compacted, often extruded from the cell. The H5 histone produced in QT6-transformed cells was found to be phosphorylated while in normal chicken fibroblasts the protein lacked this posttranslational modification. It is proposed that the chromatin-condensing role of histone H5 is inhibited by its phosphorylation.

L2 ANSWER 19 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS  
 AN 91:203428 BIOSIS  
 DN BA91:106653  
 TI MOMULV-DERIVED **SELF-INACTIVATING RETROVIRAL** VECTORS POSSESSING MULTIPLE CLONING SITES AND EXPRESSING THE RESISTANCE TO EITHER G418 OR HYGROMYCIN B.  
 AU MARTY L; POUX P; ROYER M; PIECHACZYK M  
 CS LAB. BIOLOGIE MOLECULAIRE, URA CNRS 1191, GENETIQUE MOLECULAIRE, USTL PLACE E BATAILLON 34095, MONTPELLIER, CEDEX 05.



SO BIOCHIMIE (PARIS) 72 (12). 1990. 885-888. CODEN: BICMBE ISSN: 0300-9084

LA English

AB To facilitate cloning procedures in recombinant murine leukemia virus-derived **retroviruses**, we have constructed vectors that both carry a polylinker with multiple restriction sites and express resistance to either G418 or hygromycin B. Our vectors are **self-inactivating retroviruses** that suppress interferences between LTR enhancers and internal promoters and avoid transcriptional stimulation of host cell genes. They can also be used as expression vectors in direct transfection assays, since no translation initiation codon lies between the 5' LTR and the cloning polylinker.

L2 ANSWER 20 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 90:471286 BIOSIS

DN BA90:110706

TI FUNCTIONAL ANALYSIS AND NUCLEOTIDE SEQUENCE OF THE PROMOTER REGION OF THE MURINE HCK GENE.

AU LOCK P; STANLEY E; HOLTZMANN D A; DUNN A R

CS LUDWIG INST. FOR CANCER RES., MELBOURNE TUMOUR BIOL. BRANCH, P.O. ROYAL MELBOURNE HOSP., PARKVILLE, VICTORIA 3050, AUST.

SO MOL CELL BIOL 10 (9). 1990. 4603-4611. CODEN: MCEBD4 ISSN: 0270-7306

LA English

AB The structure and function of the promoter region and exon 1 of the murine hck gene have been characterized in detail. RNase protection analysis has established that hck transcripts initiate from heterogeneous start sites located within the hck gene. Fusion gene constructs containing hck 5'-flanking sequences and the bacterial Neor gene have been introduced into the hematopoietic cell lines FDC-P1 and WEHI-265 by using a **self-inactivating retroviral** vector. The transcriptional start sites of the fusion gene are essentially identical to those of the endogenous hck gene. Analysis of infected WEHI-265 cell lines treated with bacterial lipopolysaccharide (LPS) reveals a 3- to 5-fold elevation in the levels of endogenous hck mRNA and a 1.4- to 2.6-fold increase in the level of Neor fusion gene transcripts, indicating that hck 5'-flanking sequences are capable of conferring LPS responsiveness on the Neor gene. The 5'-flanking region of the hck gene contains sequences similar to an element which is thought to be involved in the LPS responsiveness of the class II major histocompatibility gene A.alpha.k. A subset of these sequences are also found in the 5'-flanking regions of other LPS-responsive genes. Moreover, this motif is related to the consensus binding sequence of NF-kB, a transcription factor which is known to be regulated by LPS.

L2 ANSWER 21 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 90:331353 BIOSIS

DN BA90:39372

TI CONSTRUCTION AND HORMONE REGULATION OF A NOVEL **RETROVIRAL** VECTOR.

AU MEE P J; BROWN R

CS DEP. MED. ONCOL., BEATSON INST., GARSCUBE ESTATE, SWITCHBACK ROAD, BEARSDEN, GLASGOW G61 1BD, UK.

SO GENE (AMST) 88 (2). 1990. 289-292. CODEN: GENED6 ISSN: 0378-1119

LA English

AB We report the analysis of a **self-inactivating retroviral** vector, constructed to allow inducible gene expression of inserted sequences from the mouse mammary tumour virus hormonal response element. The cloning strategy has been designed to

allow for ease of insertion of the genes of interest. The vector contains the aph gene, allowing geneticin-resistance selection in mammalian cells. We have characterized dexamethasone (Dex)-induced increase in gene expression using the reporter gene encoding chloramphenicol acetyltransferase (CAT) inserted into the **retroviral** vector. We observe low basal levels of CAT activity in infected cells which is increased up to 50-fold by induction with Dex. The induction of pooled clones is 13.3-fold. Variation in Dex-induced CAT activity is observed in independently infected clones, which is not explained by proviral copy number.

L2 ANSWER 22 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS  
AN 89:428478 BIOSIS  
DN BA88:86736  
TI GERM LINE C-MYC IS NOT DOWN-REGULATED BY LOSS OR EXCLUSION OF ACTIVATING FACTORS IN MYC-INDUCED MACROPHAGE TUMORS.  
AU MANGO S E; SCHULER G D; STEELE M E R; COLE M D  
CS LEWIS THOMAS LAB., DEP. BIOL., PRINCETON UNIV., PRINCETON, N.J. 08544-1014, USA.  
SO MOL CELL BIOL 9 (8). 1989. 3482-3490. CODEN: MCEBD4 ISSN: 0270-7306  
LA English  
AB As in tumors with c-myc chromosomal translocations, c-myc **retrovirus**-induced monocyte tumors constitutively express an activated form of c-myc (the proviral gene), whereas the normal endogenous c-myc gene are transcriptionally silent. Treatment of these **retrovirus**-induced tumor cells with a number of bioactive chemicals and growth factors that are known to induce c-myc expression in cells of the monocyte lineage failed to induce the endogenous c-myc gene. In contrast, the same treatments induced the c-fos gene in both tumors and control macrophage line. To investigate c-myc suppression further, a normal copy of the human c-myc gene was introduced into tumor and control cell lines by using a **retrovirus** with **self-inactivating** long terminal repeats. This transduced normal gene was expressed at equivalent levels in all cells, regardless of the state of endogenous c-myc gene expression, and was strongly induced by agents that induce the normal gene in the control cells. The results indicate that the signal transduction pathways that normally activate the c-myc gene are functional in myc-induced tumor cells and suggest that endogenous c-myc is actively suppressed. An examination of the c-myc locus itself showed that the lack of transcriptional activity correlated with the absence of several prominent DNase I-hypersensitive sites in the 5'-flanking region of the gene but without loss of general DNase sensitivity. Furthermore, analysis of 22 methylation-sensitive restriction enzymes sites in the 5'-flanking region, first exon, and first intron indicated that the silent c-myc genes remained in the same unmethylated state as did actively expressed genes. Thus, c-myc suppression does not appear to result from the most frequently described mechanism of gene inactivation.

L2 ANSWER 23 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS  
AN 89:426460 BIOSIS  
DN BA88:84718  
TI ANALYSIS OF MAMMALIAN CELL GENETIC REGULATION IN SITU BY USING **RETROVIRUS**-DERIVED PORTABLE EXONS CARRYING THE ESCHERICHIA-COLI LAC-Z GENE.  
AU BRENNER D G; LIN-CHAO S; COHEN S N  
CS DEP. GENETICS, STANFORD UNIV. SCH. MED., STANFORD, CALIF. 94305-5120.  
SO PROC NATL ACAD SCI U S A 86 (14). 1989. 5517-5521. CODEN: PNASA6 ISSN: 0027-8424

LA English  
 AB **Self-inactivating** derivatives of Moloney murine leukemia **retrovirus** containing the Escherichia coli lacZ gene were used to detect and study the regulation of transcription initiated at chromosomally located promoters in mouse fibroblasts. The introduction of splice acceptor sites in all three translational reading frames relative to lacZ and the inclusion of an in-frame ATG translation start codon in one construct allowed synthesis of .beta.-galactosidase fusion proteins upon insertion of **retrovirus** vectors containing lacZ into introns 3' to either protein-coding or noncoding exons. Selection of lacZ-expressing cells by fluorescence-activated cell sorting and the analysis of .beta.-galactosidase production after serum deprivation has yielded lines in which lacZ was fused to genes induced by growth arrest in the G0 state.

L2 ANSWER 24 OF 24 BIOSIS COPYRIGHT 1998 BIOSIS

AN 86:319674 BIOSIS

DN BA82:43979

TI **SELF-INACTIVATING RETROVIRAL VECTORS**

DESIGNED FOR TRANSFER OF WHOLE GENES INTO MAMMALIAN CELLS.

AU YU S-F; VON RUDEN T; KANTOFF P W; GARBER C; SEIBERG M; RUTHER U;

ANDERSON W F; WAGNER E F; GILBOA E

CS DEP. MOLECULAR BIOLOGY, PRINCETON UNIV., PRINCETON, NJ 08544.

SO PROC NATL ACAD SCI U S A 83 (10). 1986. 3194-3198. CODEN: PNASAG

ISSN: 0027-8424

LA English

AB A **retrovirus**-derived vector called **self-**

**inactivating** (SIN) vector was designed for the transduction

of whole genes into mammalian cells. SIN vectors contain a deletion

of 299 base pairs in the 3' long terminal repeat (LTR), which

includes sequences encoding the enhancer and promoter functions. When

viruses derived from such vectors were used to infect NIH 3T3 cells,

the deletion was transferred to the 5' LTR, resulting in the

transcriptional inactivation of the provirus in the infected cell.

Introduction of a hybrid gene (human metallothionein-promoted c-fos)

into cells via a SIN vector was not associated with rearrangements

and led to the formation of an authentic mRNA transcript, which in

some cases was induced by cadmium. SIN vectors should be particularly

useful in gene transfer experiments designed to study the regulated

expression of genes in mammalian cells. Absence of enhancer and

promoter sequences in both LTRs of the integrated provirus should

also minimize the possibility of activating cellular oncogenes and

may provide a safer alternative to be used in human gene therapy.

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SESSION

FULL ESTIMATED COST

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24.61

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=> s promoter conversion

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      21637 PROMOTER
      17663 PROMOTERS
      28925 PROMOTER
            (PROMOTER OR PROMOTERS)
      220980 CONVERSION
      19065 CONVERSIONS
      224113 CONVERSION
            (CONVERSION OR CONVERSIONS)
L1      15 PROMOTER CONVERSION
            (PROMOTER(W) CONVERSION)

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=> s l1 and retrovir?

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      3788 RETROVIR?
L2      0 L1 AND RETROVIR?

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=> d 1 kwic

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'L2' HAS NO ANSWERS
L1      15 SEA FILE=USPAT PLU=ON  PROMOTER CONVERSION
L2      0 SEA FILE=USPAT PLU=ON  L1 AND RETROVIR?

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=> d l1 kwic

US PAT NO: 5,723,641 [IMAGE AVAILABLE] L1: 1 of 15

DETDESC:

DETD(79)

TABLE

Ex.	Promoter	Conversion %	Selectivity %
23	ZnBr.sub.2	26	83
24	ZnI.sub.2	59	82
25	ZnCl.sub.2	64	76
26	ZnSO.sub.4	31.	.

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E1	USPAT	2	GUNZ, STEFAN/IN
E2	USPAT	1	GUNZBERG, GUY W/IN
E3	USPAT	0 -->	GUNZBURG/IN
E4	USPAT	1	GUNZBURG, FRED/IN
E5	USPAT	1	GUNZEL, BODO/IN
E6	USPAT	1	GUNZEL, ERHARD P H/IN
E7	USPAT	1	GUNZEL, ERNST WERNER/IN
E8	USPAT	1	GUNZEL, HANS/IN
E9	USPAT	2	GUNZEL, PETER/IN
E10	USPAT	10	GUNZEL, RUDOLPH M JR/IN
E11	USPAT	2	GUNZELMAN, DEBORAH M/IN
E12	USPAT	1	GUNZELMANN, EDWARD J/IN

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E13	USPAT	1	GUNZELMANN, KARL HEINZ/IN
E14	USPAT	5	GUNZENHAUSER, SIGMUND/IN
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E16	USPAT	1	GUNZI, KATUHIKO/IN
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E18	USPAT	1	GUNZI, LAURENCE A/IN
E19	USPAT	1	GUNZI, THUTOMU/IN
E20	USPAT	1	GUNZIER, VOLKMAR/IN
E21	USPAT	1	GUNZINGER, ANTON/IN
E22	USPAT	1	GUNZLEN PUKALL, VOLKMAR/IN
E23	USPAT	8	GUNZLER PUKALL, VOLKMAR/IN
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=> e saller/in

E#	FILE	FREQUENCY	TERM
E1	USPAT	1	SALLENBACH, SHERRY K/IN
E2	USPAT	1	SALLENHAG, MARTIN/IN
E3	USPAT	0 -->	SALLER/IN
E4	USPAT	3	SALLER, DONALD H/IN
E5	USPAT	4	SALLER, ERIK/IN
E6	USPAT	1	SALLER, FRANZ JOHN/IN
E7	USPAT	8	SALLER, HELMUT/IN
E8	USPAT	1	SALLER, HENRY A DECEASED/IN
E9	USPAT	5	SALLER, KENNETH R/IN
E10	USPAT	1	SALLER, LOUIS RICHARD/IN
E11	USPAT	1	SALLER, MICHAEL/IN
E12	USPAT	1	SALLER, STEPHEN W/IN

=> s ell

L1            1 "SALLER, MICHAEL"/IN

=> d bib

US PAT NO: 4,738,744 [IMAGE AVAILABLE] L1: 1 of 1  
DATE ISSUED: Apr. 19, 1988  
TITLE: One-sided corrugated board machine  
INVENTOR: Michael Saller, Moosbach, Federal Republic of Germany  
ASSIGNEE: BHS-Bayerische Berg-, Hutten- und Salzwerte  
Aktiengesellschaft, Federal Republic of Germany (foreign  
corp.)  
APPL-NO: 07/097,493  
DATE FILED: Sep. 16, 1987  
ART-UNIT: 131  
PRIM-EXMR: Jerome Massie  
LEGAL-REP: Silverman, Cass, Singer & Winburn, Ltd.

=> s (homolog/ (3a) recomb?) (p) retrovir?

'HOMOLOG/ ' IS NOT A VALID FIELD CODE

=> s (homolog? (3a) recomb?) (p) retrovir?

24018 HOMOLOG?  
32980 RECOMBIN?  
3788 RETROVIR?  
L2 67 (HOMOLOG? (3A) RECOMBIN?) (P) RETROVIR?

=> d 1-10 kwic

US PAT NO: 5,721,132 [IMAGE AVAILABLE] L2: 1 of 67

DETDESC:

DETD(8)

Vectors useful for practicing the present invention include plasmids, viruses (including phage), **retroviruses**, and integratable DNA fragments (i.e., fragments integratable into the host genome by **homologous recombination**). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself.. . .

US PAT NO: 5,716,832 [IMAGE AVAILABLE] L2: 2 of 67

SUMMARY:

BSUM(7)

There . . . several shortcomings in the current use of this approach. One issue involves the generation of "live virus" (i.e., competent replicating **retrovirus**) by the producer cell line. Preparations of human therapeutics which are contaminated with **retroviruses** are not currently considered suitable for use in human therapy. For example, extreme measures are taken to exclude **retroviral** contamination of monoclonal antibodies for imaging and therapy. Live virus can in conventional producer cells when: (1) The vector genome and the helper genomes recombine with each other; (2) The vector genome or helper genome **recombines** with **homologous** cryptic endogenous **retroviral** elements in the producer cell; or (3) Cryptic endogenous **retroviral** elements reactivate (e.g., xenotropic **retroviruses** in mouse cells).

SUMMARY:



BSUM(28)

Techniques for integrating a **retroviral** genome at a specific site in the DNA of a target cell involve the use of **homologous recombination**, or alternatively, the use of a modified integrase enzyme which will recognize a specific site on the target cell genome..

DETDESC:

DETD(23)

Vector . . . and polytropic envelopes with the MoMLV gag/pol and with the vector makes these PCLs even less likely to generate replication-competent **retrovirus** by **homologous recombination** than amphotropic PCLs. Examples of the use of these methods are set forth below (see Example 2).

DETDESC:

DETD(34)

The most important safety concern for the production of **retroviral** vectors is the inherent propensity of **retroviral** PCLs to generate replication-competent **retrovirus** after introduction of a vector (Munchau et al., Virology 176:262-65, 1990). This can occur in at least two ways: 1) **homologous recombination** can occur between the therapeutic proviral DNA and the DNA encoding the MoMLV structural genes ("gag/pol" and "env") present in the PCL (discussed below under "Generation of PCLs"); and 2) generation of replication-competent virus by **homologous recombination** of the proviral DNA with the very large number of defective endogenous proviruses found in murine cells (Steffen and Weinberg, . . . Stephenson, Biochem. Biophys. Acta 458:323-54, 1976). Another safety concern with the utilization of murine cells for the production of murine **retroviral** vectors is the observation that some of the many endogenous proviral genes (**retrovirus**-like genes) in the murine genome are expressed, recognized by the **retroviral** structural gene products of murine PCLs, and delivered and expressed in target cells with an efficiency at least comparable to. . . Virol. 64:424-27, 1990). These observations strongly suggest that murine cell lines are an unsafe choice for the production of murine **retroviral** vectors for human therapeutics. To circumvent the inherent safety problems associated with murine cells, PCLs have been generated exclusively from. . .

US PAT NO: 5,716,826 [IMAGE AVAILABLE]

L2: 3 of 67

SUMMARY:

BSUM(39)

Techniques for integrating a **retroviral** genome at a specific site in the DNA of a target cell involve the use of **homologous recombination**, or alternatively, the use of a modified integrase enzyme which will recognize a specific site on the target cell genome..

DETDESC:

DETD(94)

In an eighth embodiment, the **retroviral** construct (with or without the expression of a palliative) provides a therapeutic effect by inserting itself into a virus, oncogene, or pathogenic gene, thereby inhibiting a function required for pathogenesis. This embodiment requires the direction of **retroviral** integration to a specific site in the genome by **homologous recombination**, integrase modification, or other methods (described below).

DETDESC:

DETD(250)

One technique for integrating an exogenous gene of a vector construct of a recombinant **retrovirus** into a specific site in a target cell's DNA employs **homologous recombination**. Plasmids containing sequences of DNA of greater than about 300 bp that are homologous to genomic sequences have been shown. . . .

DETDESC:

DETD(251)

In order to employ **homologous recombination** in site-specific **retroviral** integration, a vector construct should be modified such that (a) homologous sequences (to the target cell's genome) are incorporated into. . . . promoter. This approach has the effect of placing a region of homology near a potential free end of the double-stranded **retrovirus** vector genome. Free ends are known to increase the frequency of **homologous recombination** by a factor of approximately ten. In this approach, it may be necessary to defeat the normal mechanism of integration,. . . .

US PAT NO: 5,716,613 [IMAGE AVAILABLE]

L2: 4 of 67

SUMMARY:

BSUM(39)

Techniques for integrating a **retroviral** genome at a specific site in the DNA of a target cell involve the use of **homologous recombination**, or alternatively, the use of a modified integrase enzyme which will recognize a specific site on the target cell genome.. . .

DETDESC:

DETD(92)

In an eighth embodiment, the **retroviral** construct (with or without the expression of a palliative) provides a therapeutic effect by inserting itself into a virus, oncogene, or pathogenic gene, thereby inhibiting a function required for pathogenesis. This embodiment requires the direction of **retroviral** integration to a specific site in the genome by **homologous recombination**, integrase modification, or other methods (described below).

DETDESC:

DETD(248)

One technique for integrating an exogenous gene of a vector construct of a recombinant **retrovirus** into a specific site in a target cell's DNA employs **homologous recombination**. Plasmids containing sequences of DNA of greater than about 300 bp that are homologous to genomic sequences have been shown. . . .

DETDDESC:

DETD(249)

In order to employ **homologous recombination** in site-specific **retroviral** integration, a vector construct should be modified such that (a) homologous sequences (to the target cell's genome) are incorporated into. . . promoter. This approach has the effect of placing a region of homology near a potential free end of the double-stranded **retrovirus** vector genome. Free ends are known to increase the frequency of **homologous recombination** by a factor of approximately ten. In this approach, it may be necessary to defeat the normal mechanism of integration,. . . .

US PAT NO: 5,714,381 [IMAGE AVAILABLE]

L2: 5 of 67

DETDDESC:

DETD(25)

Animal . . . or mutant versions of DNA encoding a human .alpha..sub.1 adrenergic receptor or homologous animal versions of these genes, by microinjection, **retroviral** infection or other means well known to those skilled in the art, into appropriate fertilized embryos in order to produce. . . transgenic animal (Hogan B. et al. Manipulating the Mouse Embryo, A Laboratory Manual, Cold Spring Harbor Laboratory (1986)) or, 2) **Homologous recombination** (Capecchi M. R. Science 244: 1288-1292 (1989); Zimmer, A. and Gruss, P. Nature 338: 150-153 (1989)) of mutant or normal,. . . transgenic animals to alter the regulation of expression or the structure .alpha.1 of these .alpha..sub.1 adrenergic receptors. The technique of **homologous recombination** is well known in the art. It replaces the native gene with the inserted gene and so is useful for. . . .

US PAT NO: 5,714,313 [IMAGE AVAILABLE]

L2: 6 of 67

DETDDESC:

DETD(23)

The present invention further provides a vector comprising a selectable marker gene inserted into a retrotransposon, wherein the retrotransposon comprises a **retroviral** reverse transcriptase/RNase H gene domain and wherein the selectable marker gene contains an intron inserted into a coding sequence of. . . thus provides an assay for detecting inhibitors of certain retrotranspositional events. For example, inhibitors of any of the activities of **retroviral** reverse transcriptase can be detected. Additionally, inhibitors of the Ty protease or any other substituted protease, can be detected, as can inhibitors of cellular genes involved in **homologous recombination** events. Once a lead compound has been identified by the present method, its specific mode of inhibition can be determined. . . method,

however, provides an inexpensive, easy assay to screen compounds initially. Further, because of the precision with which the selected **retroviral** RT/RH can be substituted into the present vector, compounds very specific for inhibiting the selected RT/RH can be detected.

DETDESC:

DETD(27)

The His.sup.+ events obtained with HART21 and any present vector substituting different **retroviral** RT/RH domains into the retrotransposon can result, instead of from retrotransposition, from a related process termed cDNA-mediated **recombination**, a **homologous recombination** process. cDNA-mediated recombination predominates when reverse transcription products do not undergo transpositional integration catalyzed by the Tyl-encoded enzyme integrase. The cDNA product undergoes **homologous recombination** with endogenous Ty elements residing in the yeast genome. The His.sup.+ cDNA recombination events are completely dependent on functional RT/RH and, therefore, serve as a faithful biological indicator of **retroviral** RT/RH activities in yeast (FIG. 5). Additionally, a His.sup.+ event requires the RT to act as an RNA-dependent DNA polymerase. . . the present assay to detect inhibitors of RT/RH provides a useful screen for compounds for drug intervention in therapy against **retroviral** infection.

DETDESC:

DETD(28)

That a **homologous recombination** event, rather than a retrotransposition event, occurs with any of the present **retroviral** RT/RH vectors to produce a His.sup.+ phenotype can be tested by any of several standard methods, several of which are. . . result in a significant amount of insertion of the reverse transcription product into the plasmid DNA in the cell, whereas **homologous recombination** results in insertion of the product into both the chromosome and the plasmid by virtue of the presence of Ty elements in both. Therefore, the marker on the reverse transcription product, if **homologous recombination** occurs, becomes genetically linked with markers on the plasmid, which can be readily detected by standard means.

US PAT NO: 5,712,148 [IMAGE AVAILABLE]

L2: 7 of 67

DETDESC:

DETD(26)

Animal . . . of normal or mutant versions of DNA encoding a mammalian transporter or homologous animal versions of these genes, by microinjection, **retroviral** infection or other means well known to those skilled in the art, into appropriate fertilized embryos in order to produce a transgenic animal (Hogan et al., 1986) **Homologous recombination** (Cappechi, M. R., 1989; Zimmer A., and Gruss, P., 1989) of mutant or normal, human or animal versions of these. . . gene locus in transgenic animals to alter the regulation of expression or the structure of these transporters. The technique of **homologous recombination** is well known in the art. It replaces the native gene with the inserted gene and so is useful for. . .

US PAT NO: 5,705,732 [IMAGE AVAILABLE]

L2: 8 of 67

DETDESC:

DETD(72)

Once cloned, the .beta..sub.2 M gene is subcloned into a plasmid based or preferentially a **retroviral**-based vector (the "gene targeting vector") such that the reading frame of the .beta..sub.2 M gene is disrupted by insertion of. . . additional selection gene (the "negative selection gene"), outside of the disrupted .beta..sub.2 m gene region which allows for selection against non-**homologous recombination**, i.e., for selection against incorporation of the entire plasmid into the genetic information of the cell rather than just the. . .

US PAT NO: 5,698,446 [IMAGE AVAILABLE]

L2: 9 of 67

DETDESC:

DETD(22)

Since the various nucleotide sequences required for generating therapeutically useful **retroviral** vectors are typically harbored on more than one nucleic acid molecule and frequently contain homologous regions between the vectors, consequently, recombination may occur between the nucleotide sequences of **retroviral** gag/pol and env and other nucleic acid sequences that are conserved among the various vectors present in the packaging and/or. . . acid cassette may be inserted 3' of the homologous region in the amphotropic env expression vector CMVenvAm(Dra), so that a **homologous recombination** event will result in the production of a nucleic acid molecule containing both the 5' inhibitor/5' splice and the 3'. . .

US PAT NO: 5,691,177 [IMAGE AVAILABLE]

L2: 10 of 67

SUMMARY:

BSUM(39)

Techniques for integrating a **retroviral** genome at a specific site in the DNA of a target cell involve the use of **homologous recombination**, or alternatively, the use of a modified integrase enzyme which will recognize a specific site on the target cell genome.. . .

DETDESC:

DETD(86)

In an eighth embodiment, the **retroviral** construct (with or without the expression of a palliative) provides a therapeutic effect by inserting itself into a virus, oncogene, or pathogenic gene, thereby inhibiting a function required for pathogenesis. This embodiment requires the direction of **retroviral** integration to a specific site in the genome by **homologous recombination**, integrase modification, or other methods (described below).

DETDESC:

DETD(242)

One technique for integrating an exogenous gene of a vector construct of a recombinant **retrovirus** into a specific site in a target cell's DNA employs **homologous recombination**. Plasmids containing sequences of DNA of greater than about 300 bp that are homologous to genomic sequences have been shown. . . .

DETD(243)

DETD(243)

In order to employ **homologous recombination** in site-specific **retroviral** integration, a vector construct should be modified such that (a) homologous sequences (to the target cell's genome) are incorporated into. . . promoter. This approach has the effect of placing a region of homology near a potential free end of the double-stranded **retrovirus** vector genome. Free ends are known to increase the frequency of **homologous recombination** by a factor of approximately ten. In this approach, it may be necessary to defeat the normal mechanism of integration,. . . .

=> d 11-20 kwic

US PAT NO: 5,686,573 [IMAGE AVAILABLE]

L2: 11 of 67

DETD(32)

DETD(32)

Vectors useful for practicing the present invention include plasmids, viruses (including phage), **retroviruses**, and integrable DNA fragments (i.e., fragments integrable into the host genome by **homologous recombination**). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself.. . .

DETD(38)

DETD(38)

Cloned . . . of the present invention are useful in gene therapy, for replacing defective dopamine receptor genes in vivo. For such purposes, **retroviral** vectors as described in U.S. Pat. No. 4,650,764 to Temin and Watanabe or U.S. Pat. No. 4,861,719 to Miller may. . . be employed. Cloned genes of the present invention, or fragments thereof, may also be used in gene therapy carried out **homologous recombination** or site-directed mutagenesis. See generally Smithies et al., 1985, Nature 917: 230-234; Thomas and Capecchi, 1987, Cell 51: 503-512; Bertling,. . . .

US PAT NO: 5,683,912 [IMAGE AVAILABLE]

L2: 12 of 67

DETD(58)

DETD(58)

The transfer of nucleic acid material into mammalian hosts for the purpose of generating transgenic animals can be accomplished by

microinjection, **retroviral** infection or other means well known to those skilled in the art, of the material into appropriate fertilized embryos. (See, for example, Hogan et al., Manipulating the Mouse Embryo: A Laboratory Manual (Cold Spring Harbor Laboratory, 1986). **Homologous recombination** can also be used for the generation of transgenic animals according to the present invention. **Homologous recombination** techniques are well known in the art. **Homologous recombination** replaces a native (endogenous) gene with a recombinant or mutated gene to produce an animal that cannot express a native (endogenous) alpha9 receptor subunit but can express, for example, a mutated receptor subunit. In contrast to **homologous recombination**, microinjection adds genes to the host genome, without removing host genes. Microinjection can produce a transgenic animal that is capable. . .

US PAT NO: 5,681,746 [IMAGE AVAILABLE]

L2: 13 of 67

DETDESC:

DETD(42)

An . . . packaging cell lines useful in the invention is the production therefrom of replication incompetent virions, or avoidance of generating replication-competent **retrovirus** (RCR) [Munchau et al., Virology, vol. 176:262-65, 1990]. This will ensure that infectious **retroviral** particles harboring the recombinant retroviral vectors of the invention will be incapable of independent replication in target cells, be they. . . the possibility of insertional mutagenesis and its associated problems. RCR production can occur in at least two ways: (1) through **homologous recombination** between the therapeutic proviral DNA and the DNA encoding the **retroviral** structural genes ("gag/pol" and "env") present in the packaging cell line; and (2) generation of replication-competent virus by **homologous recombination** of the proviral DNA with the very large number of defective endogenous proviruses found in the packaging cell line.

DETDESC:

DETD(61)

In . . . leads to integration of the viral genome into a chromosome of the recipient cell, as occurs in the case of **retroviral** infection, **homologous recombination** or use of a modified integrase enzyme which directs insertion to a specific site can be utilized. Such site-specific insertion. . .

US PAT NO: 5,677,139 [IMAGE AVAILABLE]

L2: 14 of 67

SUMMARY:

BSUM(43)

Many . . . include vital vectors, episomal plasmid vectors, stably integrating plasmid vectors, and artificial chromosome vectors. Vital vectors include those derived from **retroviruses**, adenoviruses, adeno-associated viruses, herpesviruses, and pox viruses. Vital vectors may be delivered as virus particles or by another delivery mechanism. . . including synthetic nucleic acids. Vectors may be designed to integrate at a specific location in the genome by insertion or **homologous recombination**, may integrate randomly or may remain in

the nucleus as a stable episomal nucleic acid. All of the above vectors.

US PAT NO: 5,675,063 [IMAGE AVAILABLE]

L2: 15 of 67

SUMMARY:

BSUM(26)

DNA can be introduced into embryonic stem (ES) cells by transfection, **retroviral** infection, or electroporation. The most important advantage for gene transfer into animals is that cells carrying the transgene can be selected for before being injected into a blastocyst. For example, ES cells were infected with **retroviral** vectors, or transfected with plasmids, carrying the neo gene. This gene confers resistance to the antibiotic G418. Only ES cells. . . F2 generation were G418-resistant. Because ES cells can be manipulated in vitro before injection into the embryo, geneticists can use **homologous recombination** to produce transgenic animals with mutations, specific genes or to replace a mutant gene with the normal equivalent.

US PAT NO: 5,665,585 [IMAGE AVAILABLE]

L2: 16 of 67

DETDESC:

DETD(37)

In . . . or, in a most preferred embodiment, be assisted by transformation with a vector inserting it into the host chromosome by **homologous recombination**, for example, with **retroviral** vectors, transposons or other DNA elements which promote integration of DNA sequences in chromosomes. A vector is employed which is. . . T. reesei, such vectors may provide a gene encoded by T. reesei, such as the cellobiohydrolase I gene, to promote **homologous recombination** at a specific site on the host chromosome.

US PAT NO: 5,658,786 [IMAGE AVAILABLE]

L2: 17 of 67

DETDESC:

DETD(26)

Animal . . . of normal or mutant versions of DNA encoding a mammalian transporter or homologous animal versions of these genes, by microinjection, **retroviral** infection or other means well known to those skilled in the art, into appropriate fertilized embryos in order to produce a transgenic animal (24) or 2) **Homologous recombination** (7,82) of mutant or normal, human or animal versions of these genes with the native gene locus in transgenic animals to alter the regulation of expression or the structure of these transporters. The technique of **homologous recombination** is well known in the art. It replaces the native gene with the inserted gene and so is useful for. . .

US PAT NO: 5,658,783 [IMAGE AVAILABLE]

L2: 18 of 67

DETDESC:

DETD(8)

Vectors useful for practicing the present invention include plasmids,



viruses (including phage), **retroviruses**, and integratable DNA fragments (i.e., fragments integratable into the host genome by **homologous recombination**). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself.. . .

US PAT NO: 5,658,782 [IMAGE AVAILABLE]

L2: 19 of 67

DETDESC:

DETD(11)

Vectors useful for practicing the present invention include plasmids, viruses (including phage), **retroviruses**, and integratable DNA fragments (i.e., fragments integratable into the host genome by **homologous recombination**). The vector replicates and functions independently of the host genome, or may, in some instances, integrate into the genome itself.. . .

US PAT NO: 5,656,465 [IMAGE AVAILABLE]

L2: 20 of 67

DETDESC:

DETD(27)

Recombinant fowlpox viruses (FPV) were constructed by **homologous recombination** in a manner analogous to that previously described (Jenkins, et al. AIDS Research and Human **Retroviruses** 7:991-998 (1991). Foreign sequences were inserted at the Bg/II site in the BamHI J region of the FPV genome. FPV. . .

=> d 2,3,4 bib ab

US PAT NO: 5,716,832 [IMAGE AVAILABLE]

L2: 2 of 67

DATE ISSUED: Feb. 10, 1998

TITLE: Packaging cells

INVENTOR: Jack R. Barber, San Diego, CA  
Douglas J. Jolly, La Jolla, CA  
James G. Respass, San Diego, CA  
Stephen M. W. Chang, San Diego, CA

ASSIGNEE: Chiron Viagene, Inc. (U.S. corp.)

APPL-NO: 08/462,492

DATE FILED: Jun. 5, 1995

ART-UNIT: 185

PRIM-EXMR: Mindy Fleisher

ASST-EXMR: Robert Schwartzman

LEGAL-REP: Norman J. Kruse, Robert P. Blackburn

US PAT NO: 5,716,832 [IMAGE AVAILABLE]

L2: 2 of 67

ABSTRACT:

The invention described herein allows the production of recombinant retroviruses (retroviral vector particles) from producer cells which are safer and of higher titre than normal. In addition, methods are provided for making helper cells which, when a recombinant retrovirus genome is introduced to make a producer line, produce particles that are targeted toward particular cell types. Methods are also provided for making recombinant retrovirus systems adapted to infect a particular cell type,

such as a tumor, by binding the retrovirus or recombinant retrovirus in the particular cell type. Methods are also provided for producing recombinant retroviruses which integrate in a specific small number of places in the host genome, and for producing recombinant retroviruses from transgenic animals.

US PAT NO: 5,716,826 [IMAGE AVAILABLE] L2: 3 of 67  
DATE ISSUED: Feb. 10, 1998  
TITLE: Recombinant retroviruses  
INVENTOR: Harry E. Gruber, Rancho Santa Fe, CA  
Douglas J. Jolly, Leucadia, CA  
James G. Respass, San Diego, CA  
Paul K. Laikind, San Diego, CA  
ASSIGNEE: Chiron Viagene, Inc. (U.S. corp.)  
APPL-NO: 08/136,739  
DATE FILED: Oct. 12, 1993  
ART-UNIT: 185  
PRIM-EXMR: George G. Elliott  
ASST-EXMR: Robert Schwartzman  
LEGAL-REP: Norman J. Seed & Berry Kruse, Robert P. Blackburn

US PAT NO: 5,716,826 [IMAGE AVAILABLE] L2: 3 of 67

ABSTRACT:

Recombinant retroviruses carrying a vector construct capable of preventing, inhibiting, stabilizing or reversing infectious, cancerous or auto-immune diseases are disclosed. More specifically, the recombinant retroviruses of the present invention are useful for (a) stimulating a specific immune response to an antigen or a pathogenic antigen; (b) inhibiting a function of a pathogenic agent, such as a virus; and (c) inhibiting the interaction of an agent with a host cell receptor. In addition, eucaryotic cells infected with, and pharmaceutical compositions containing such a recombinant retrovirus are disclosed. Various methods for producing recombinant retroviruses having unique characteristics, and methods for producing transgenic packaging animals or insects are also disclosed.

US PAT NO: 5,716,613 [IMAGE AVAILABLE] L2: 4 of 67  
DATE ISSUED: Feb. 10, 1998  
TITLE: Recombinant retroviruses  
INVENTOR: Harry E. Guber, San Diego, CA  
Douglas J. Jolly, La Jolla, CA  
James G. Respass, San Diego, CA  
Paul K. Laikind, San Diego, CA  
ASSIGNEE: Chiron Viagene, Inc. (U.S. corp.)  
APPL-NO: 08/474,736  
DATE FILED: Jun. 7, 1995  
ART-UNIT: 185  
PRIM-EXMR: George G. Elliott  
ASST-EXMR: Robert Schwartzman  
LEGAL-REP: Norman J. Seed & Berry Kruse, Robert P. Blackburn

US PAT NO: 5,716,613 [IMAGE AVAILABLE] L2: 4 of 67

ABSTRACT:

Recombinant retroviruses carrying a vector construct capable of preventing, inhibiting, stabilizing or reversing infectious, cancerous or auto-immune diseases are disclosed. More specifically, the recombinant

retroviruses of the present invention are useful for (a) stimulating a specific immune response to an antigen or a pathogenic antigen; (b) inhibiting a function of a pathogenic agent, such as a virus; and (c) inhibiting the interaction of an agent with a host cell receptor. In addition, eucaryotic cells infected with, and pharmaceutical compositions containing such a recombinant retrovirus are disclosed. Various methods for producing recombinant retroviruses having unique characteristics, and methods for producing transgenic packaging animals or insects are also disclosed.

=> d 2 clms

US PAT NO: 5,716,832 [IMAGE AVAILABLE]

L2: 2 of 67

CLAIMS:

CLMS(1)

We claim:

1. A method of producing a recombinant retrovirus, comprising growing a producer cell having a genome comprising:

- (a) a gene of interest and a packaging signal of a first retrovirus;
- (b) gag and pol genes of the first retrovirus, absent a packaging signal;
- (c) a hybrid env gone absent a packaging signal, the product of said hybrid env gone comprising a cytoplasmic segment of the first retrovirus, and a binding segment exogenous to the first retrovirus.

CLMS(2)

2. A xenotropic packaging cell line which, upon introduction of a vector construct, produces viral particles uncontaminated by replication competent virus.

CLMS(3)

3. The packaging cell line of claim 2 wherein the cell line produces at least equal vector titre as compared to a standard mouse amphotropic packaging cell line PA 317 when HT1080 cells are infected.

CLMS(4)

4. A polytropic packaging cell line which, upon introduction of a vector construct, produces viral particles uncontaminated by replication competent virus.

CLMS(5)

5. The packaging cell line of claim 4 wherein the packaging cell line, upon introduction of a vector construct, produces at least a ten-fold increase in vector titre as compared to a standard measure amphotropic packaging cell line PA 317 when 293 cells are infected.

CLMS(6)

6. A polytropic packaging cell line wherein the packaging cell line, upon introduction of a vector construct, produces viral particles which infect cells of kidney lineage, but not cells or fibroblast, epithelial,

T-cell or monocyte lineage.

CLMS(7)

7. A non-mouse packaging cell the carrying on separate operons the genes for gag/pol and env, said operons lacking retroviral LTR sequences and which, upon introduction of an N2 type vector construct, produces no helper virus after at least twenty days passage in culture.

CLMS(8)

8. The cell line of claim 7 wherein the cell line is an amphotropic packaging cell line.

CLMS(9)

9. The cell line of claim 7 wherein the cell line is a polytropic packaging cell line.

CLMS(10)

10. The cell line of claim 7 wherein the cell line is a xenotropic packaging cell line.

CLMS(11)

11. A method of producing a recombinant retrovirus, comprising:
- (a) introducing packaging genes from a retroviral vector system into a cell line, said cell line having no endogenous proviruses which produce transcripts packageable by the retroviral vector system;
  - (b) selecting for cells that produce at least a ten-fold increase in viral packaging protein as compared to a standard mouse amphotropic packaging cell line PA317, and that, upon introduction of a vector construct, produce at least a ten-fold increase in vector titre as compared to a standard mouse amphotropic packaging cell line PA317; and
  - (c) growing the cells selected in step (b) such that recombinant retrovirus is produced.

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=> e gunzburg w h/au

E1	1	GUNZBURG T S/AU
E2	2	GUNZBURG W/AU
E3	38 -->	GUNZBURG W H/AU
E4	3	GUNZBURGER D/AU
E5	5	GUNZBURGER L/AU
E6	14	GUNZBURGER L K/AU
E7	3	GUNZCLER P/AU
E8	1	GUNZE C M B/AU
E9	1	GUNZEL/AU
E10	1	GUNZEL A/AU
E11	13	GUNZEL A R/AU
E12	6	GUNZEL APEL A R/AU













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L1 38 "GUNZBURG W H"/AU

=> e saller r m/au

E1	1	SALLER L/AU
E2	68	SALLER R/AU
E3	9 -->	SALLER R M/AU
E4	2	SALLER S/AU
E5	7	SALLER U/AU
E6	2	SALLER Y/AU
E7	1	SALLERA SANMARTI L/AU
E8	2	SALLERAS J M/AU
E9	8	SALLERAS J P/AU
E10	114	SALLERAS L/AU
E11	2	SALLERAS L I/AU
E12	2	SALLERAS L Y/AU

=> s e3

L2 9 "SALLER R M"/AU

=> s l1 and l2

L3 3 L1 AND L2

=> d 1-3 bib ab

L3 ANSWER 1 OF 3 MEDLINE  
AN 1998105826 MEDLINE  
DN 98105826  
TI Construction and characterization of a hybrid mouse mammary tumor virus/murine leukemia virus-based retroviral vector.  
AU **Saller R M**; Ozturk F; Salmons B; **Gunzburg W H**  
CS Institut fur Molekulare Virologie, GSF-Forschungszentrum fur Umwelt und Gesundheit, Oberschleissheim, Germany.  
SO JOURNAL OF VIROLOGY, (1998 Feb) 72 (2) 1699-703.  
Journal code: KCV. ISSN: 0022-538X.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 199804  
EW 19980403  
AB Mouse mammary tumor virus (MMTV)-based vectors are characterized by low titers. In an effort to transfer MMTV-specific regulation of gene expression to a more efficient murine leukemia virus (MLV) vector, we have replaced the complete 3' U3 region of MLV with the complete U3 region of MMTV. Virus titers were not significantly affected by this modification, there was no impairment of reverse transcription and integration, and after infection of cells, the MMTV promoter is duplicated and translocated to the 5' long terminal repeat, resulting in glucocorticoid-regulatable RNA expression.

L3 ANSWER 2 OF 3 MEDLINE  
AN 96022220 MEDLINE  
DN 96022220

TI Construction of retroviral vectors for targeted delivery and expression of therapeutic genes.  
 AU Salmons B; **Saller R M**; Baumann J G; **Gunzburg W H**  
 CS Bavarian Nordic Research Institute, Oberschleissheim, Germany..  
 SO LEUKEMIA, (1995 Oct) 9 Suppl 1 S53-60. Ref: 44  
 Journal code: LEU. ISSN: 0887-6924.  
 CY ENGLAND: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 General Review; (REVIEW)  
 (REVIEW, TUTORIAL)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199602  
 AB Current gene therapy protocols take an ex vivo approach in which cells are removed from a patient, genetically modified and then reimplanted. However this kind of approach is both cumbersome and costly, requiring high tech facilities and is limited to cell types that can be easily cultured. The in vivo delivery of genes by retroviral vectors will greatly facilitate gene therapy protocols of the future. However before in vivo gene therapy becomes a reality a number of problems must be overcome. Ideally therapeutic genes should be delivered only to the relevant cell type and/or expressed in this cell type. Strategies are described that (I) limit therapeutic gene delivery, using pseudotyping or vectors based on retroviruses that show a restricted infection spectrum or (II) limit the expression of transferred genes by inclusion of tissue specific promoters or cis acting regulatory elements. The combination of some of these strategies should permit the construction of novel retroviral vectors that provide safe and targeted in vivo gene transfer.

L3 ANSWER 3 OF 3 MEDLINE  
 AN 95344681 MEDLINE  
 DN 95344681  
 TI Retroviral vectors directed to predefined cell types for gene therapy.  
 AU **Gunzburg W H**; **Saller R M**; Salmons B  
 CS GSF-Forschungszentrum fur Umwelt und Gesundheit GmbH, Institut fur Molekulare Virologie, Oberschleissheim, Germany..  
 SO BIOLOGICALS, (1995 Mar) 23 (1) 5-12. Ref: 41  
 Journal code: AMW. ISSN: 1045-1056.  
 CY ENGLAND: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 General Review; (REVIEW)  
 (REVIEW, TUTORIAL)  
 LA English  
 FS Priority Journals  
 EM 199511

=> s (homolog? (3a) recomb?) and retrovir?

L4 167 (HOMOLOG? (3A) RECOMB?) AND RETROVIR?

=> s homologous recombination and retrovir?

L5 132 HOMOLOGOUS RECOMBINATION AND RETROVIR?

=> s 15 range=,1995

L6 101 L5

=> s 15 range=,1994

L7 89 L5

=> duplicate remove 17

DUPLICATE PREFERENCE IS 'MEDLINE, BIOSIS'  
KEEP DUPLICATES FROM MORE THAN ONE FILE? Y/(N):n

PROCESSING COMPLETED FOR L7

L8 53 DUPLICATE REMOVE L7 (36 DUPLICATES REMOVED)

=> d 1-10 bib ab

L8 ANSWER 1 OF 53 MEDLINE DUPLICATE 1  
AN 94187082 MEDLINE  
DN 94187082  
TI **Retrovirus** recombination depends on the length of sequence  
identity and is not error prone.  
AU Zhang J; Temin H M  
CS McArdle Laboratory for Cancer Research, University of  
Wisconsin-Madison 53706.  
NC CA-22443 (NCI)  
CA-07175 (NCI)  
SO JOURNAL OF VIROLOGY, (1994 Apr) 68 (4) 2409-14.  
Journal code: KCV. ISSN: 0022-538X.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 199406  
AB **Retroviruses**, as a result of the presence of two identical  
genomic RNA molecules in their virions, recombine at a high rate.  
When nonhomologous RNA is present in the dimer RNA molecules,  
nonhomologous recombination can occur, although the rate is very  
low, only 0.1% of the rate of essentially **homologous**  
**recombination** (J. Zhang and H. M. Temin, Science  
259:234-238, 1993). We found, as is found in naturally occurring  
highly oncogenic **retroviruses** (J. Zhang and H. M. Temin,  
J. Virol. 67:1747-1751, 1993), that the crossovers usually occur at  
a short region of sequence identity. We modified the previously  
studied vectors to study the effect of different lengths of short  
regions of sequence identity in the midst of otherwise nonidentical  
sequences. We found that the efficiency of recombination depends on  
the length of this sequence identity. However, the highest rate in  
such molecules remained lower than for recombination between  
essentially homologous molecules, even when there was extensive  
sequence identity. Junction sequences of the recombinants indicated  
that **retrovirus** recombination is not an error-prone  
process as was reported for human immunodeficiency virus reverse  
transcriptase by using a cell-free system (J. A. Peliska and S. J.  
Benkovic, Science 258:1112-1118, 1992).

L8 ANSWER 2 OF 53 MEDLINE  
AN 94331440 MEDLINE  
DN 94331440  
TI Characterization of a replication-competent **retrovirus**  
resulting from recombination of packaging and vector sequences.

AU Otto E; Jones-Trower A; Vanin E F; Stambaugh K; Mueller S N;  
 Anderson W F; McGarrity G J  
 CS Genetic Therapy Inc., Gaithersburg, MD 20878.  
 SO HUMAN GENE THERAPY, (1994 May) 5 (5) 567-75.  
 Journal code: A12. ISSN: 1043-0342.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199411  
 AB A replication-competent **retrovirus** (RCR) was detected by  
 S+/L- assays in three lots of **retroviral** vector G1Na that  
 were harvested on consecutive days from a single culture of  
 PA317/G1Na producer cells. Using a number of **retrovirus**  
 -specific primer pairs, it was shown that this RCR was a novel  
 recombinant created by exchanges between G1Na and helper sequence  
 pPAM3 and was not an existing RCR introduced by cross-contamination.  
 Sequencing of clones of DNA amplified in six independent PCR  
 reactions confirmed that the 3' portion of this RCR was composed of  
**retroviral** envelope sequences unique to pPAM3 joined to a 3'  
 long terminal repeat (LTR) unique to G1Na. Comparison of pPAM3 and  
 G1Na sequences at the site corresponding to this junction revealed a  
 short segment of patchy nucleotide identity (8 out of 10 bp),  
 suggesting that these helper and vector sequences were joined by  
**homologous recombination**. Generation of RCR by  
 exchanges between helper and vector sequences underscores the  
 necessity of testing by efficient methods all **retroviral**  
 vectors for the presence of RCR before their use. Production of 171  
 lots (855 liters) of various **retroviral** vectors that were  
 free of RCR, including 42 lots of G1Na, however, indicates that the  
 combination of exchanges required to generate an RCR are infrequent  
 in this system.

L8 ANSWER 3 OF 53 MEDLINE DUPLICATE 2  
 AN 94076409 MEDLINE  
 DN 94076409

TI One **retroviral** RNA is sufficient for synthesis of viral  
 DNA.

AU Jones J S; Allan R W; Temin H M  
 CS McArdle Laboratory for Cancer Research, University of Wisconsin,  
 Madison 53706.  
 NC CA22443 (NCI)  
 CA07175 (NCI)  
 NCI CA09075 (NCI)  
 SO JOURNAL OF VIROLOGY, (1994 Jan) 68 (1) 207-16.  
 Journal code: KCV. ISSN: 0022-538X.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199403  
 AB We used previously characterized spleen necrosis virus-based  
**retroviral** vectors and helper cells to study the strand  
 transfers that occur during the reverse-transcription phase of a  
 single cycle of **retroviral** replication. The conditions  
 used selected only for formation of an active provirus rather than  
 for expression of multiple drug resistance markers. In  
 nonrecombinant proviruses the minus- and plus-strand DNA primer  
 transfers were almost completely intramolecular. However, as  
 previously reported, recombinant proviruses contained approximately



equal proportions of inter- and intramolecular minus-strand DNA primer transfers. Thus, we conclude that in the absence of recombination, one molecule of **retroviral** RNA is sufficient for viral DNA synthesis. Large deletions and deletions with insertions were detected primarily at a limited number of positions which appear to be hot spots for such events, the primer binding site and regions containing multiple inverted repeats. At these hot spots, the rate of deletions and deletions with insertions visible with PCR was about 10% per genome per replication cycle. Other deletions and deletions with insertions (detectable with PCR) occurred at a rate of about 0.5%/kb per replication cycle. Crossovers occurred at a rate of about 6%/kb per replication cycle under single-selection conditions. This rate is comparable to the rate that we reported previously under double-selection conditions, indicating that **retroviral homologous recombination** is not highly error prone. The combined rates of deletions and deletions with insertions at hot spots (10% per genome per replication cycle) and other sites (0.5%/kb per replication cycle) and the rate of crossovers (6%/kb per replication cycle) indicate that on average, full-size (10-kb) type C **retroviruses** undergo an additional or aberrant strand transfer about once per cycle of infection.

L8 ANSWER 4 OF 53 MEDLINE  
AN 94082449 MEDLINE  
DN 94082449  
TI Pathogenicity of a subgroup C feline leukemia virus (FeLV) is augmented when administered in association with certain FeLV recombinants.  
AU Mathes L E; Pandey R; Chakrabarti R; Hofman F M; Hayes K A; Stromberg P; Roy-Burman P  
CS Center for Retrovirus Research, Ohio State University, Columbus 43210.  
NC CA51485 (NCI)  
T32-CA09320 (NCI)  
SO VIROLOGY, (1994 Jan) 198 (1) 185-95.  
Journal code: XEA. ISSN: 0042-6822.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 199403  
AB There is evidence to suggest that infectious feline leukemia viruses (FeLVs) may be altered biologically because of **homologous recombination** with non-infectious endogenous FeLV (enFeLV) sequences in the infected cells. To evaluate the role of such recombination events in FeLV pathogenesis, a molecular clone of subgroup C FeLV, Sarma strain (FSC), was tested for induction of aplastic anemia in the absence or presence of mixtures of recombinants between FSC and an enFeLV element. In the recombinants, FSC sequences in the viral surface glycoprotein (SU) protein were variably replaced by the corresponding sequences of the enFeLV. The results showed that the virus mixtures varied in their infectivity to neonatal specific pathogen-free cats. One group of mixtures, although exhibiting relatively reduced infectivity, represented the most acute disease-inducing agents. The presence of recombinants in this mixture significantly accelerated the development of erythrocyte aplasia compared to cats infected with FSC alone. In addition, infected cells appeared to be distributed differently in various hematopoietic organs with respect to infection with FSC

versus viral mixture. Viral recombinants which were present in this inoculum mixture, however, could not be detected in the plasma or infected tissues of the cats at the end stage of the disease, although their presence in the plasma at the early stages could be detected. Clearly, parental FSC outgrew the recombinants in the infected animals, since its detection was prominent at all stages of the progression of the disease. Therefore, we hypothesize that recombinants initially present in the infected animals, while only poorly replicated compared to FSC in the host, might have had the opportunity to infect certain target cells (potentially erythroid progenitor cells) and then disappeared with the associated cytopathic effect.

L8 ANSWER 5 OF 53 BIOSIS COPYRIGHT 1998 BIOSIS  
 AN 94:482569 BIOSIS  
 DN 97495569  
 TI Versatile **retroviral** vectors for potential use in gene therapy.  
 AU Hawley R G; Lieu F H L; Fong A Z C; Hawley T S  
 CS Div. Cancer Res., Reichmann Res. Build., S218, Sunnybrook Health Sci. Cent., 2075 Bayview Ave., Toronto, ON M4N 3M5, CAN  
 SO Gene Therapy 1 (2). 1994. 136-138.  
 LA English  
 AB A set of **retroviral** vectors is described whose capacity for high efficiency transduction of functional genes into undifferentiated murine embryonic and haematopoietic cells makes them ideally suited for preclinical studies with murine models. Multiple unique cloning sites permit insertion of genes into the vectors such that no selectable marker exists or either the neomycin phosphotransferase (neo) gene, the hygromycin B phosphotransferase (hph) gene or the puromycin N-acetyl transferase (pac) gene is included as a dominantly acting selectable marker. Because the sequences in the viral gag region shown to improve the encapsidation of viral RNA have been modified to prevent viral protein synthesis and all env sequences have been removed to eliminate helper virus production by **homologous recombination** with packaging DNA, these vectors might prove useful in human gene therapy protocols.

L8 ANSWER 6 OF 53 MEDLINE  
 AN 94265646 MEDLINE  
 DN 94265646  
 TI [Transgenic mice--biological fundamentals, practices and applications].  
 Transgene Mause--Biologische Grundlagen, Praktiken und Anwendung.  
 AU Balling R  
 CS Institut fur Saugetiergenetik, GSF-Forschungszentrum fur Umwelt und Gesundheit, Oberschleissheim..  
 SO DTW. DEUTSCHE TIERARZTLICHE WOCHENSCHRIFT, (1994 Mar) 101 (3) 94-5.  
 Ref: 10  
 Journal code: ECT. ISSN: 0341-6593.  
 CY GERMANY: Germany, Federal Republic of  
 DT Journal; Article; (JOURNAL ARTICLE)  
 General Review; (REVIEW)  
 (REVIEW, TUTORIAL)  
 LA German  
 EM 199409  
 AB The transfer of genes into the germline of mice has become a standard technique of modern biomedical research. In addition to direct microinjection of DNA into the pronuclei of fertilized mouse

eggs, genes can now also be inactivated via **homologous recombination** in embryonic stem cells (gene targeting). In the future it will be possible to inactivate any cloned gene via **homologous recombination** in ES-cells and establish corresponding mouse mutants.

L8 ANSWER 7 OF 53 BIOSIS COPYRIGHT 1998 BIOSIS  
AN 93:585044 BIOSIS  
DN 97004414  
TI High rate of genetic rearrangement during replication of a Moloney murine leukemia virus-based vector.  
AU Varela-Echavarria A; Proppock C M; Ron Y; Dougherty J P  
CS Dep. Mol. Genetics Microbiol., Robert Wood Johnson Med. Sch., University Medicine, Dentistry New Jersey, 675 Hoes Lane, Piscataway, NJ 08854-5635, USA  
SO Journal of Virology 67 (11). 1993. 6357-6364. ISSN: 0022-538X  
LA English  
AB A protocol was designed to measure the forward mutation rate over an entire gene replicated as part of a Moloney murine leukemia virus-based vector. For these studies, the herpes simplex virus thymidine kinase (tk) gene under the control of the spleen necrosis virus U3 promoter was used as target sequence since it allows selection for either the functional or the inactivated gene. Our results indicate that after one round of **retroviral** replication, the tk gene is inactivated at an average rate of 0.08 per cycle of replication. Southern blotting revealed that the majority of the mutant proviruses resulted from gross rearrangements and that deletions of spleen necrosis virus and tk sequences were the most frequent cause of the gene inactivation. Sequence analysis of the mutant proviruses suggested that homologous as well as nonhomologous recombination was involved in the observed rearrangements. Some mutations consisted of simple deletions, and others consisted of deletions combined with insertions. The frequency at which these mutations occurred during one cycle of **retroviral** replication provides evidence indicating that Moloney murine leukemia virus-based vectors may undergo genetic rearrangement at high rates. The high rate of rearrangement and its relevance for **retrovirus**-mediated gene transfer are discussed.

L8 ANSWER 8 OF 53 MEDLINE  
AN 93267761 MEDLINE  
DN 93267761  
TI Alteration of location of dimer linkage sequence in **retroviral** RNA: little effect on replication or **homologous recombination**.  
AU Jones J S; Allan R W; Temin H M  
CS McArdle Laboratory for Cancer Research, University of Wisconsin, Madison 53706.  
NC CA22443 (NCI)  
CA07175 (NCI)  
CA09075 (NCI)  
SO JOURNAL OF VIROLOGY, (1993 Jun) 67 (6) 3151-8.  
Journal code: KCV. ISSN: 0022-538X.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 199308  
AB **Retrovirus** particles contain a dimer of **retroviral**

genomic RNA. A defined region of the **retrovirus** genome has previously been shown to be important for both dimerization and encapsidation. To study the importance of the position of this encapsidation and dimerization signal for **retroviral** replication and **homologous recombination**, we used a previously described spleen necrosis virus-based helper cell system. This system allows **retroviral** vectors with multiple genetic markers to be studied after a single cycle of **retroviral** replication. The sequence responsible for dimerization, the encapsidation/dimer linkage sequence (E/DLS), was moved from its normal location near the 5' end of the **retroviral** genome to a location near the 3' end of the genome. We characterized four pairs of **retroviral** vectors: (i) with both E/DLSs at the 5' ends of the genomes, (ii) with both E/DLSs at the 3' ends of the genomes, and (iii) two with one E/DLS at the 5' end of the genome and one at the 3' end of the genome. We found that moving the E/DLS to the 3' end of the genome resulted in at most an approximately factor of 5 reduction in virus titer in a single cycle of **retroviral** replication. Furthermore, we found no changes that were attributable to the alteration of the position of the E/DLS in the minus-strand DNA primer transfers or the plus-strand DNA primer transfers, the rate of **homologous recombination**, or the number of internal template switches in recombinant proviruses. These results indicate that any alignment or conformation necessary for **retroviral** replication or recombination is not the result of the position of the E/DLS.

L8 ANSWER 9 OF 53 MEDLINE DUPLICATE 4  
 AN 93390966 MEDLINE  
 DN 93390966  
 TI Long cellular repeats flanking a defective HTLV-I provirus: implication for site-targeted integration.  
 AU Kubota S; Furuta R; Maki M; Siomi H; Hatanaka M  
 CS Institute for Virus Research, Kyoto University, Japan..  
 SO ONCOGENE, (1993 Oct) 8 (10) 2873-7.  
 Journal code: ONC. ISSN: 0950-9232.  
 CY ENGLAND: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199312  
 AB **Retroviruses** generally integrate as proviruses which are flanked by long-terminal repeats (LTRs) on both 5' and 3' ends. Since these LTRs are required for the efficient integration mediated by the viral integrase, it is believed that defective proviruses with a single LTR are normally formed by deletion after integration. However, we found no deletion of cellular sequences around the integration site of such a defective HTLV-1. Rather, we identified 99 bp-long direct repeats adjacent to both ends of the defective provirus. The repeated cellular sequences contained a potential poly(A) signal followed by a **retroviral** primer-binding-site-like sequence. The presence of the direct repeats of cellular sequences can be explained by the integration of the defective virus through **homologous recombination** between cellular and viral read-through sequences.

L8 ANSWER 10 OF 53 MEDLINE DUPLICATE 5  
 AN 93345780 MEDLINE  
 DN 93345780

TI Synthetic retrotransposon vectors for gene therapy.  
 AU Chakraborty A K; Zink M A; Boman B M; Hodgson C P  
 CS Creighton Cancer Center/Dept. of Biomedical Sciences, Creighton  
 School of Medicine, Omaha, Nebraska 68178..  
 NC 1R29GM41314-4 (NIGMS)  
 SO FASEB JOURNAL, (1993 Jul) 7 (10) 971-7.  
 Journal code: FAS. ISSN: 0892-6638.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199311  
 AB New gene therapy methods are rapidly being developed to permit the  
 expression of tumor suppressor genes, cytotoxins, anticancer  
 antigens, and immunoregulatory proteins in the treatment of cancer.  
 Large-scale testing in humans has been delayed by questions  
 concerning the safety and effectiveness of preferred  
**retroviral** vectors and helper cells. These vector systems  
 are limited by their ability to undergo **homologous**  
**recombination** with endogenous **retroviruses** or  
 helper-viral sequences, resulting in release of replication-  
 competent **retrovirus** (RCR). In addition, transcriptional  
 inactivation of the **retroviral** promoter often occurs,  
 caused in part by methylation of CpG islands in the  
**retroviral** long terminal repeats (LTRs). We report the  
 production of highly specific retrovectors using gene amplification  
 together with oligonucleotide building blocks. The synthetic vectors  
 were based on mouse VL30 retrotransposon NVL3, and lacked homology  
 to **retroviral** helper gene sequences. Three of four  
 constructs made by gene amplification yielded biologically active  
 vectors. These constructs efficiently transmitted and stably  
 inserted a neomycin resistance marker gene into the genome of  
 recipient cells, expressing an abundant RNA species of the expected  
 size in the absence of detectable replication competent  
**retrovirus**. The vectors and techniques described enable  
 widely applicable expression modes using generic helper cells, and  
 require only approximately 1.3 kb of cis-acting vector RNA sequences  
 for faithful transfer and expression of genetic material.

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FILE 'MEDLINE, BIOSIS' ENTERED AT 13:17:34 ON 12 MAR 1998

E GUNZBURG W H/AU  
 L1 38 S E3  
 E SALLER R M/AU  
 L2 9 S E3  
 L3 3 S L1 AND L2  
 L4 167 S (HOMOLOG? (3A) RECOMB?) AND RETROVIR?  
 L5 132 S HOMOLOGOUS RECOMBINATION AND RETROVIR?  
 L6 101 S L5  
 L7 89 S L5  
 L8 53 DUPLICATE REMOVE L7 (36 DUPLICATES REMOVED)

=> s 15 and (host or chromosom?)

L9 36 L5 AND (HOST OR CHROMOSOM?)

=> d 1-10 bib ab

L9 ANSWER 1 OF 36 MEDLINE  
AN 1998105778 MEDLINE  
DN 98105778  
TI The application of a **homologous recombination**  
assay revealed amino acid residues in an LTR-retrotransposon that  
were critical for integration.  
AU Atwood A; Choi J; Levin H L  
CS Laboratory of Eukaryotic Gene Regulation, National Institute of  
Child Health and Human Development, National Institutes of Health,  
Bethesda, Maryland 20892, USA.  
SO JOURNAL OF VIROLOGY, (1998 Feb) 72 (2) 1324-33.  
Journal code: KCV. ISSN: 0022-538X.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 199804  
EW 19980403  
AB **Retroviruses** and their relatives, the LTR-  
retrotransposons, possess an integrase protein (IN) that is required  
for the insertion of reverse transcripts into the genome of  
**host** cells. *Schizosaccharomyces pombe* is the **host**  
of Tfl, an LTR-retrotransposon with integration activity that can be  
studied by using techniques of yeast genetics. In this study, we  
sought to identify amino acid substitutions in Tfl that specifically  
affected the integration step of transposition. In addition to  
seeking amino acid substitutions in IN, we also explored the  
possibility that other Tfl proteins contributed to integration. By  
comparing the results of genetic assays that monitored both  
transposition and reverse transcription, we were able to seek point  
mutations throughout Tfl that blocked transposition but not the  
synthesis of reverse transcripts. These mutant versions of Tfl were  
candidates of elements that possessed defects in the integration  
step of transposition. Five mutations in Tfl that resulted in low  
levels of integration were found to be located in the IN protein:  
two substitutions in the N-terminal Zn domain, two in the catalytic  
core, and one in the C-terminal domain. These results suggested that  
each of the three IN domains was required for Tfl transposition. The  
potential role of these five amino acid residues in the function of  
IN is discussed. Two of the mutations that reduced integration  
mapped to the RNase H (RH) domain of Tfl reverse transcriptase. The  
Tfl elements with the RH mutations produced high levels of reverse  
transcripts, as determined by recombination and DNA blot analysis.  
These results indicated that the RH of Tfl possesses a function  
critical for transposition that is independent of the accumulation  
of reverse transcripts.

L9 ANSWER 2 OF 36 MEDLINE  
AN 95336307 MEDLINE  
DN 95336307  
TI LTR-directed **homologous recombination** of  
full-length HIV-1 provirus clone in recA(-) bacteria.  
AU Yamada K; Morozumi H; Okamoto T  
CS Department of Molecular Genetics, Nagoya City University Medical  
School, Aichi, Japan.  
SO ARCHIVES OF VIROLOGY, (1995) 140 (6) 1007-14.  
Journal code: 8L7. ISSN: 0304-8608.

CY Austria  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199510  
 AB During molecular cloning of full-length **retroviral** plasmid clones occurrence of **homologous recombination** (HR) between LTR regions is frequently observed. In order to evaluate appropriate **host** bacterial strains for cloning such HR-prone plasmids, we utilized a linearized template plasmid containing a full-length HIV-1 proviral sequence. The plasmid was linearized within the viral sequence so that plasmid transformed bacteria would grow only when the plasmid was circularized by HR. Using this genetic system for detecting HR, we evaluated the frequency of HR in various recA(-) bacterial strains which are commercially available and in some recA-null strains in which recA-defective phenotype was constructed by P1 transduction. We found that HR occurred even in recA-null strains although in lesser frequencies. The nucleotide sequence analysis at the junction of recombination revealed no loss, insertion or duplication of DNA sequence. It is suggested that recombination machinery other than the RecA system is involved.

L9 ANSWER 3 OF 36 MEDLINE  
 AN 95211017 MEDLINE  
 DN 95211017  
 TI Activities of granulocyte-macrophage colony-stimulating factor revealed by gene transfer and gene knockout studies.  
 AU Dranoff G; Mulligan R C  
 CS Dana-Farber Cancer Institute, Boston, Massachusetts.  
 SO STEM CELLS, (1994) 12 Suppl 1 173-82; discussion 182-4. Ref: 65  
 Journal code: BN2. ISSN: 1066-5099.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 General Review; (REVIEW)  
 (REVIEW, TUTORIAL)  
 LA English  
 FS Priority Journals  
 EM 199507  
 AB We used **retroviral** mediated gene transfer and gene knockout technologies to explore the in vivo functions of murine granulocyte-macrophage colony-stimulating factor (GM-CSF) [1, 2]. In tumor vaccination experiments, GM-CSF was the most potent molecule of a large number of cytokines, adhesion molecules and other immunomodulators for the induction of specific and long-lasting anti-tumor immunity. Vaccination required activities of both CD4 and CD8 positive lymphocytes, and likely involved the augmentation by GM-CSF of **host** professional antigen-presenting cell function. Mice engineered by **homologous recombination** techniques in embryonic stem cells to lack GM-CSF demonstrated no significant perturbations in steady-state hematopoiesis. All mutant animals, however, developed the accumulation of surfactant proteins and lipids in the alveolar space, the defining feature of the idiopathic human disorder pulmonary alveolar proteinosis. Surfactant lipid and protein content were increased in the absence of alterations in surfactant protein mRNA, supporting the concept that surfactant clearance or catabolism was perturbed. Extensive lymphoid hyperplasia associated with lung airways and blood vessels was also found, yet no infectious agents could be isolated. These results demonstrate that GM-CSF is not an

essential growth factor for basal hematopoiesis and reveal an unexpected, critical role for GM-CSF in pulmonary homeostasis. It is tempting to speculate that the ability of GM-CSF to modulate the uptake and processing of particulate material underlies the mechanisms of immunostimulation and surfactant accumulation.

L9 ANSWER 4 OF 36 MEDLINE  
AN 95133154 MEDLINE  
DN 95133154  
TI Intrachromosomal recombination mediated by the polyomavirus large T antigen.  
AU Laurent S; Frances V; Bastin M  
CS Department of Biochemistry, University of Sherbrooke, Quebec, Canada..  
SO VIROLOGY, (1995 Jan 10) 206 (1) 227-33.  
Journal code: XEA. ISSN: 0042-6822.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 199504  
AB We used a spleen necrosis virus-based **retroviral** vector to introduce the polyomavirus replication origin into rat cells and developed a system to analyze **homologous recombination** events that do not reconstitute a selectable marker. Introduction of the gene coding for the polyomavirus large T antigen into the cell lines by DNA transfection promoted high-frequency recombination between the two **retroviral** LTRs, leading to amplification and excision of DNA sequences. To analyze homology requirements, we constructed cell lines carrying only the replication origin without exogenous repeats. Most of the cell lines sustained high-frequency recombination, presumably by undergoing **homologous recombination** between repetitive DNA lying in the vicinity of the integrated origin. Our results indicate that **homologous recombination** promoted by large T antigen does not require recombination hot spots in the viral genome other than the replication origin and they explain the cytotoxicity observed in some cell types when large T antigen is expressed in the presence of a functional origin.

L9 ANSWER 5 OF 36 MEDLINE  
AN 95129544 MEDLINE  
DN 95129544  
TI Concerted evolution of the tandem array encoding primate U2 snRNA occurs in situ, without changing the cytological context of the RNU2 locus.  
AU Pavelitz T; Rusche L; Matera A G; Scharf J M; Weiner A M  
CS Department of Molecular Biophysics and Biochemistry, Yale University School of Medicine, New Haven, CT 06510.  
NC GM31073 (NIGMS)  
GM41624 (NIGMS)  
SO EMBO JOURNAL, (1995 Jan 3) 14 (1) 169-77.  
Journal code: EMB. ISSN: 0261-4189.  
CY ENGLAND: United Kingdom  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals  
OS GENBANK-L37793; GENBANK-U57614  
EM 199504  
AB In primates, the tandemly repeated genes encoding U2 small nuclear



RNA evolve concertedly, i.e. the sequence of the U2 repeat unit is essentially homogeneous within each species but differs somewhat between species. Using **chromosome** painting and the NGFR gene as an outside marker, we show that the U2 tandem array (RNU2) has remained at the same **chromosomal** locus (equivalent to human 17q21) through multiple speciation events over > 35 million years leading to the Old World monkey and hominoid lineages. The data suggest that the U2 tandem repeat, once established in the primate lineage, contained sequence elements favoring perpetuation and concerted evolution of the array in situ, despite a pericentric inversion in chimpanzee, a reciprocal translocation in gorilla and a paracentric inversion in orang utan. Comparison of the 11 kb U2 repeat unit found in baboon and other Old World monkeys with the 6 kb U2 repeat unit in humans and other hominids revealed that an ancestral U2 repeat unit was expanded by insertion of a 5 kb **retrovirus** bearing 1 kb long terminal repeats (LTRs). Subsequent excision of the provirus by **homologous recombination** between the LTRs generated a 6 kb U2 repeat unit containing a solo LTR. Remarkably, both junctions between the human U2 tandem array and flanking **chromosomal** DNA at 17q21 fall within the solo LTR sequence, suggesting a role for the LTR in the origin or maintenance of the primate U2 array.

L9 ANSWER 6 OF 36 MEDLINE

AN 94331440 MEDLINE

DN 94331440

TI Characterization of a replication-competent **retrovirus** resulting from recombination of packaging and vector sequences.

AU Otto E; Jones-Trower A; Vanin E F; Stambaugh K; Mueller S N; Anderson W F; McGarrity G J

CS Genetic Therapy Inc., Gaithersburg, MD 20878.

SO HUMAN GENE THERAPY, (1994 May) 5 (5) 567-75.

Journal code: A12. ISSN: 1043-0342.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 199411

AB A replication-competent **retrovirus** (RCR) was detected by S+/L- assays in three lots of **retroviral** vector G1Na that were harvested on consecutive days from a single culture of PA317/G1Na producer cells. Using a number of **retrovirus**-specific primer pairs, it was shown that this RCR was a novel recombinant created by exchanges between G1Na and helper sequence pPAM3 and was not an existing RCR introduced by cross-contamination. Sequencing of clones of DNA amplified in six independent PCR reactions confirmed that the 3' portion of this RCR was composed of **retroviral** envelope sequences unique to pPAM3 joined to a 3' long terminal repeat (LTR) unique to G1Na. Comparison of pPAM3 and G1Na sequences at the site corresponding to this junction revealed a short segment of patchy nucleotide identity (8 out of 10 bp), suggesting that these helper and vector sequences were joined by **homologous recombination**. Generation of RCR by exchanges between helper and vector sequences underscores the necessity of testing by efficient methods all **retroviral** vectors for the presence of RCR before their use. Production of 171 lots (855 liters) of various **retroviral** vectors that were free of RCR, including 42 lots of G1Na, however, indicates that the combination of exchanges required to generate an RCR are infrequent in this system.

L9 ANSWER 7 OF 36 MEDLINE  
 AN 94082449 MEDLINE  
 DN 94082449  
 TI Pathogenicity of a subgroup C feline leukemia virus (FeLV) is augmented when administered in association with certain FeLV recombinants.  
 AU Mathes L E; Pandey R; Chakrabarti R; Hofman F M; Hayes K A; Stromberg P; Roy-Burman P  
 CS Center for Retrovirus Research, Ohio State University, Columbus 43210.  
 NC CA51485 (NCI)  
 T32-CA09320 (NCI)  
 SO VIROLOGY, (1994 Jan) 198 (1) 185-95.  
 Journal code: XEA. ISSN: 0042-6822.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals; Cancer Journals  
 EM 199403  
 AB There is evidence to suggest that infectious feline leukemia viruses (FeLVs) may be altered biologically because of **homologous recombination** with non-infectious endogenous FeLV (enFeLV) sequences in the infected cells. To evaluate the role of such recombination events in FeLV pathogenesis, a molecular clone of subgroup C FeLV, Sarma strain (FSC), was tested for induction of aplastic anemia in the absence or presence of mixtures of recombinants between FSC and an enFeLV element. In the recombinants, FSC sequences in the viral surface glycoprotein (SU) protein were variably replaced by the corresponding sequences of the enFeLV. The results showed that the virus mixtures varied in their infectivity to neonatal specific pathogen-free cats. One group of mixtures, although exhibiting relatively reduced infectivity, represented the most acute disease-inducing agents. The presence of recombinants in this mixture significantly accelerated the development of erythrocyte aplasia compared to cats infected with FSC alone. In addition, infected cells appeared to be distributed differently in various hematopoietic organs with respect to infection with FSC versus viral mixture. Viral recombinants which were present in this inoculum mixture, however, could not be detected in the plasma or infected tissues of the cats at the end stage of the disease, although their presence in the plasma at the early stages could be detected. Clearly, parental FSC outgrew the recombinants in the infected animals, since its detection was prominent at all stages of the progression of the disease. Therefore, we hypothesize that recombinants initially present in the infected animals, while only poorly replicated compared to FSC in the **host**, might have had the opportunity to infect certain target cells (potentially erythroid progenitor cells) and then disappeared with the associated cytopathic effect.

L9 ANSWER 8 OF 36 MEDLINE  
 AN 93267761 MEDLINE  
 DN 93267761  
 TI Alteration of location of dimer linkage sequence in **retroviral** RNA: little effect on replication or **homologous recombination**.  
 AU Jones J S; Allan R W; Temin H M  
 CS McArdle Laboratory for Cancer Research, University of Wisconsin, Madison 53706.

NC CA22443 (NCI)  
CA07175 (NCI)  
CA09075 (NCI)  
SO JOURNAL OF VIROLOGY, (1993 Jun) 67 (6) 3151-8.  
Journal code: KCV. ISSN: 0022-538X.  
CY United States  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals; Cancer Journals  
EM 1993  
AB **Retrovirus** particles contain a dimer of **retroviral** genome RNA. A defined region of the **retrovirus** genome has previously been shown to be important for both dimerization and encapsidation. To study the importance of the position of this encapsidation and dimerization signal for **retroviral** replication and **homologous recombination**, we used a previously described spleen necrosis virus-based helper cell system. This system allows **retroviral** vectors with multiple genetic markers to be studied after a single cycle of **retroviral** replication. The sequence responsible for dimerization, the encapsidation/dimer linkage sequence (E/DLS), was moved from its normal location near the 5' end of the **retroviral** genome to a location near the 3' end of the genome. We characterized four pairs of **retroviral** vectors: (i) with both E/DLSs at the 5' ends of the genomes, (ii) with both E/DLSs at the 3' ends of the genomes, and (iii) two with one E/DLS at the 5' end of the genome and one at the 3' end of the genome. We found that moving the E/DLS to the 3' end of the genome resulted in at most an approximately factor of 5 reduction in virus titer in a single cycle of **retroviral** replication. Furthermore, we found no changes that were attributable to the alteration of the position of the E/DLS in the minus-strand DNA primer transfers or the plus-strand DNA primer transfers, the rate of **homologous recombination**, or the number of internal template switches in recombinant proviruses. These results indicate that any alignment or conformation necessary for **retroviral** replication or recombination is not the result of the position of the E/DLS.

L9 ANSWER 9 OF 36 MEDLINE  
AN 93254440 MEDLINE  
DN 93254440  
TI [Complete nucleotide sequence of Rous sarcoma virus variants adapted to duck cells].  
Polnaya nukleotidnaia posledovatel'nost' adaptirovannogo k kletkam utok varianta virusa sarkomy rausa.  
AU Kashura V I; Kavsan V M; Ryndich A V; Lazurkevich Z V; Zubak S V; Popov A V; Dostalova V; Glozhanek I  
SO MOLECULARNAIA BIOLOGIIA, (1993 Mar-Apr) 27 (2) 436-50.  
Journal code: NGX. ISSN: 0026-8984.  
CY RUSSIAN Russian Federation  
DT Journal; Article; (JOURNAL ARTICLE)  
LA Russian  
FS Priority Journals  
EM 1993  
AB Subject: C avian sarcoma viruses efficiently infect and transform but do not efficiently replicate in duck cells. Nucleotide sequence analysis of Prague strain of Rous sarcoma virus adapted by numerous passages on duck embryonic fibroblasts (daPr-RSV-C) showed that adaptation of originally chicken virus to duck cells correlated with changes in viral genome, first of all in gp85-coding domain of env-gene.

Besides changes in LTR and src-gene sequences could play a role in widening of host range for this virus. The major changes of d. RSV-C in comparison with original Pr-RSV-C appeared to be the result of **homologous recombinations** with corresponding regions of chicken endogenous **retroviruses**.

L9 ANSWER 10 OF 36 MEDLINE  
 AN 9207 MEDLINE  
 DN 9207  
 TI The dispersion of defective endogenous murine **retroviral** elements suggests retrotransposition-mediated amplification.  
 AU Fredham M; Policastro P F; Wilson M C  
 CS Scripps Research Institute, Department of Molecular Biology and Neuropharmacology, La Jolla, CA 92037.  
 NC CA 31 (NCI)  
 CA 0 (NCI)  
 SO DNA CELL BIOLOGY, (1991 Dec) 10 (10) 713-22.  
 JOURN CODE: AF9. ISSN: 1044-5498.  
 CY United States  
 DT Journal Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 1992  
 AB The dispersion of four replication-defective endogenous proviruses, originally detected in 129 strain mice and shown to have extensive deletions of gag, pol, and env gene regions, was investigated in 13 inbred strains and substrains of mice. Using probes to sequences flanking the integration sites in 129 mice, unique genomic Eco RI fragments were assigned to each of the four endogenous proviral elements. Analyses revealed that certain of these proviral elements are present both in strains closely related to strain 129 (i.e., strains 101 and LP/J) and in more distantly related strains (i.e., strains BALB/cJ, A/J, and C3H/HeJ). In mouse strains lacking proviral integration at a particular locus, the size of the corresponding Eco RI genomic fragment and absence of a characteristic Kpn I site indicated the lack of a residual solitary long terminal repeat. Hybridization of oligonucleotide probes that distinguish the specific deletions present within these elements identified additional analogous proviral integrations at many different sites in all strains investigated. These data indicate that the diversification of these proviral elements found in inbred strains is generated by integration of new copies, rather than excision through **homologous recombination**. Moreover, the results are consistent with other endogenous **retroviruses** providing the trans-acting proteins necessary to package the defective viral RNA.

=> d 11-2 11-2 ab

L9 ANSWER 11 OF 36 MEDLINE  
 AN 9007 MEDLINE  
 DN 9007  
 TI **Homologous recombination** between the LTRs of a human **retrovirus**-like element causes a 5-kb deletion in two siblings.  
 AU Magee J; Goodchild N L  
 CS Terry Fox Laboratory, British Columbia Cancer Research Centre, University of British Columbia, Vancouver, Canada..  
 SO AMER JOURNAL OF HUMAN GENETICS, (1989 Dec) 45 (6) 848-54.

Jour Code: 3IM. ISSN: 0002-9297.  
 CY Unit States  
 DT Journal Article; (JOURNAL ARTICLE)  
 LA Engl  
 FS Price Journals  
 EM 1990  
 AB The RTVL-H family of human endogenous **retrovirus**-like elements consists of approximately 1,000 intact members and of a similar number of solitary long terminal repeats (LTRs). In this study the genetic heterogeneity of these elements has been investigated using unique flanking probes isolated from cDNA clones containing RTVL-H sequences. Four such probes were used to screen a panel of human DNA samples for genetic differences. One of these probes detected a 5.0-kb deletion in two related individuals. Cloning and DNA hybridization analysis indicated that the nondeleted common allele contained an intact RTVL-H element, whereas the deleted variant allele contained only a single LTR. DNA sequence comparisons strongly suggest that the deletion is due to **homologous recombination** between the 5' and 3' LTRs of the RTVL-H sequence. This is the first reported case of a DNA deletion in humans that is due to an LTR-LTR excision event.

L9 ANSWER 2 OF 36 MEDLINE  
 AN 8938 MEDLINE  
 DN 8938  
 TI Prospects for correction of thalassemia by genetic engineering.  
 AU Gale  
 CS Department of Medicine, UCLA School of Medicine 90024.  
 NC CA23 (NCI)  
 SO PROCEEDINGS IN CLINICAL AND BIOLOGICAL RESEARCH, (1989) 309 141-59.  
 Ref:

Jour Code: PZ5. ISSN: 0361-7742.  
 CY Unit States  
 DT Journal Article; (JOURNAL ARTICLE)  
 Gene Review; (REVIEW)  
 (REVIEW TUTORIAL)  
 LA Engl  
 FS Price Journals  
 EM 1989  
 AB The **thalassemias** are diverse genetic disorders characterized by abnormal synthesis rates of one or more proteins constituting hemoglobin (globin-chains). In the beta-thalassemias, genes encoding the beta-globin chain are intact but are abnormally transcribed or, less often, translated. In the alpha-thalassemias, genes encoding the alpha-globin chain are often deleted; abnormal transcription can also occur. The human beta- and alpha-globin genes were molecularly cloned. This review considers attempts to introduce these genes in mammalian cells by physical techniques such as **chromosome** transfer, transfection, fusion, micro-injection, electroporation and **homologous recombination** or by using DNA or RNA viruses such as **retroviruses**. Currently, inefficient gene expression in host cells and the need for precise cognate regulation of globin gene expression are the major limitations to applying genetic engineering to thalassemia.

L9 ANSWER 2 OF 36 MEDLINE  
 AN 8920 MEDLINE  
 DN 8920  
 TI Gene targeting with **retroviral** vectors: recombination by gene conversion into regions of nonhomology.

AU Ellis Bernstein A  
 CS Department of Medical Genetics, University of Toronto, Ontario,  
 Canada  
 SO MOLECULAR AND CELLULAR BIOLOGY, (1989 Apr) 9 (4) 1621-7.  
 JOURNAL CODE: NGY. ISSN: 0270-7306.  
 CY Unit States  
 DT Journal Article; (JOURNAL ARTICLE)  
 LA English  
 FS Price Journals  
 EM 1989  
 AB We have designed and constructed integration-defective  
 retroviral vectors to explore their potential for gene  
 targeting in mammalian cells. Two nonoverlapping deletion mutants of  
 the bacterial neomycin resistance (neo) gene were used to detect  
 homologous recombination events between viral and  
 chromosomal sequences. Stable neo gene correction events  
 were detected at a frequency of approximately 1 G418r cell per 3 x  
 10<sup>6</sup> infected cells. Analysis of the functional neo gene in  
 independent targeted cell clones indicated that unintegrated  
 retroviral linear DNA recombined with the target by gene  
 conversion for variable distances into regions of nonhomology. In  
 addition, transient neo gene correction events which were associated  
 with complete loss of the chromosomal target sequences  
 were observed. These results demonstrated that retroviral  
 vectors can recombine with homologous chromosomal  
 sequences in rodent and human cells.

L9 ANSWER OF 36 MEDLINE  
 AN 8925 MEDLINE  
 DN 8925  
 TI Activation of the c-H-ras proto-oncogene by retrovirus  
 insertion and chromosomal rearrangement in a Moloney  
 leukemia virus-induced T-cell leukemia.  
 AU Ihle J; Smith-White B; Sisson B; Parker D; Blair D G; Schultz A;  
 Koza J; Lunsford R D; Askew D; Weinstein Y; et al  
 CS Molecular Mechanisms of Carcinogenesis Laboratory, National Cancer  
 Institute-Frederick, Cancer Research Facility, Maryland 21701.  
 NC NO1-101 (NCI)  
 CA21 (NCI)  
 SO JOURNAL OF VIROLOGY, (1989 Jul) 63 (7) 2959-66.  
 JOURNAL CODE: KCV. ISSN: 0022-538X.  
 CY Unit States  
 DT Journal Article; (JOURNAL ARTICLE)  
 LA English  
 FS Price Journals; Cancer Journals  
 OS GENETICS 127193  
 EM 1989  
 AB A rearrangement of the c-H-ras locus was detected in a T-cell line  
 (DA-2) established from a Moloney leukemia virus-induced tumor. This  
 rearrangement was associated with the high-level expression of H-ras  
 RNA, the H-ras gene product, p21. DNA from DA-2 cells transformed  
 fibroblasts in DNA transfection experiments, and the transformed  
 fibroblasts contained the rearranged H-ras locus. The rearrangement  
 involved one allele and was present in tissue from the primary tumor  
 from which the cell line was isolated. Cloning and sequencing of the  
 rearranged allele and comparison with the normal allele demonstrated  
 that the rearrangement was complex and probably resulted from the  
 insertion of a retrovirus in the H-ras locus between a  
 5' coding exon and the first coding exon and a subsequent  
 homologous recombination between this provirus and

L9	ANSL	OF 36 MEDLINE
AN	881.	MEDLINE
DN	881.	
TI	Rec	tion between two integrated proviruses, one of which was
	ins	near c-myc in a <b>retrovirus</b> -induced rat thymoma:
	imp	ons for tumor progression.
AU	Laz	Tsichlis P N
CS	Fox	Cancer Center, Philadelphia, Pennsylvania 19111.
NC	CA-	(NCI)
	CA-	(NCI)
	RR-	(NCRR)
SO	JOU	OF VIROLOGY, (1988 Mar) 62 (3) 788-94.
	Jou	code: KCV. ISSN: 0022-538X.
CY	Uni	ates
DT	Jou	Article; (JOURNAL ARTICLE)
LA	Eng	
FS	Pri	Journals; Cancer Journals
OS	GEN	9096
EM	198	
AB	Of	oney murine leukemia virus (MoMuLV)-induced rat thymomas, 2
	con	rearrangement. in c-myc. In one of these tumors the
	obse	rearrangement was not due to the insertion of an intact
	MoM	ovirus. The rearranged c-myc DNA fragment from this thymoma
	was	and examined by restriction endonuclease mapping,
	hyb	tion to MoMuLV proviral DNA probes, and DNA sequence
	ana	These analyses revealed that the c-myc rearrangement in
	this	was due to the presence of a partially duplicated MoMuLV
	long	inal repeat (LTR) 5' to c-myc exon 1. The orientation of
	this	structure was opposite to the transcriptional orientation
	of c	The sequences at the 3' flanking side of the LTR structure
	were	from a cellular DNA region which maps to the same
	chrom	one as c-myc (chromosome 7), although to a
	site	ant from this proto-oncogene. These findings present
	evic	for a homologous <b>recombination</b> event
	occ	between sequences of two proviruses integrated on the same
	chrom	one, one of which was inserted near the c-myc
	prot	ogene. The recombination product contains three copies of
	the	LTR 72-base-pair direct repeat and is associated with a
	high	of c-myc expression. The reciprocal product of this
	reco	tion was not detected. We propose that recombination
	betw	ologous sequences may play a significant role in the
	gene	of chromosomal rearrangements and therefore in
	tum	uction and progression.

L9	ANS:	OF 36 MEDLINE
AN	852	MEDLINE
DN	852:	
TI	Struc	and biological activity of human homologs of the raf/mil
	onc	
AU	Bon:	; Kerby S B; Stravre P; Gunnell M A; Mark G; Rapp U R
SO	MOL:	AND CELLULAR BIOLOGY, (1985 Jun) 5 (6) 1400-7.

Jou de: NGY. ISSN: 0270-7306.  
 CY Uni tes  
 DT Jou Article; (JOURNAL ARTICLE)  
 LA Eng  
 FS Pri Journals  
 OS GEN 11376; GENBANK-L00213; GENBANK-L00206; GENBANK-L00207;  
 GEN 10208; GENBANK-L00209; GENBANK-L00210; GENBANK-L00211;  
 GEN 100212; GENBANK-M11377  
 EM 1985  
 AB Two genes homologous to the raf/mil oncogene have been cloned and sequenced. One, c-raf-2, is a processed pseudogene; the other, c-raf-1, contains nine exons homologous to both raf and mil and two additional exons homologous to mil. A 3' portion of c-raf-1 contains six of the seven amino acid differences relative to murine raf and can substitute for the 3' portion of v-raf in a transfection assay. Sequence homologies between c-raf-1 and Moloney leukemia virus at both ends of v-raf indicate that the viral gene was acquired by **homologous recombination**. Although the data are consistent with the traditional model of **retrovirus** transduction, they also raise the possibility that transduction occurred in a double crossover event between proviral DNA and the murine gene.

L9 ANS OF 36 MEDLINE  
 AN 851 MEDLINE  
 DN 851  
 TI Recombination between a defective **retrovirus** and homologous sequences in **host** DNA: reversion by patch replication  
 AU Schaffhausen B S; Berg P; Colicelli J; Goff S P  
 NC CA (NCI)  
 SO JOURNAL OF VIROLOGY, (1985 Mar) 53 (3) 719-26.  
 Jou de: KCV. ISSN: 0022-538X.  
 CY Uni tes  
 DT Jou Article; (JOURNAL ARTICLE)  
 LA Eng  
 FS Pri Journals; Cancer Journals  
 OS GEN 13364; GENBANK-M17738  
 EM 1985  
 AB The genomes of mammalian species contain multiple copies of sequences homologous to exogenous **retroviruses**. When a mutant **retrovirus** carrying a lethal deletion in an essential viral gene was introduced into mammalian cells, revertant viruses appeared and spread throughout the culture. Analysis of one such revertant showed that the mutation had been repaired by **homologous recombination** with endogenous sequences. Our results suggest that defective **retroviruses** can depend upon the genetic complement of the **host** cell to replicate and recombine in viral genes.

L9 ANS OF 36 MEDLINE  
 AN 850 MEDLINE  
 DN 850  
 TI Nucleotide sequences of the oncogene v-rel in Rous sarcoma virus strain T and its cellular homolog, the proto-oncogene c-rel.  
 AU Wilentz Z; K C; Eggleston K; Temin H M  
 NC CA (NCI)  
 CA (NCI)  
 CA (NCI)



SO JOURNAL OF VIROLOGY, (1984 Oct) 52 (1) 172-82.  
 Jou Title: KCV. ISSN: 0022-538X.  
 CY Uni Titles  
 DT Journal Article; (JOURNAL ARTICLE)  
 LA Eng  
 FS Pri Journals; Cancer Journals  
 OS GENBANK-K00537; GENBANK-K00555; GENBANK-K02447; GENBANK-K02448;  
 GENBANK-K02449; GENBANK-K02450; GENBANK-K02451; GENBANK-K02452;  
 GENBANK-K02453; GENBANK-K02454; GENBANK-K02455  
 EM 198  
 AB Ret Endotheliosis virus strain T (Rev-T) is a highly oncogenic  
 rep non-defective **retrovirus** which contains the  
 onc c-rel. It is thought that Rev-T arose when a virus similar  
 to the helper virus of Rev-T, infected a turkey and  
 rec with c-rel from that turkey. There is one large c-rel  
 loc the turkey genome which contains all of the sequences  
 hom to v-rel (K. C. Wilhelmsen and H. M. Temin, J. Virol.  
 49: 1984). We have sequenced v-rel and its flanking  
 seq each of the regions of the c-rel locus from turkey that  
 are homologous to v-rel and their flanking sequences, and the coding  
 seq for env and part of pol of Rev-A. The v-rel coding  
 seq can be translated into a 503-amino acid  
 env out-of-frame-env fusion polypeptide. We have not detected  
 any clones in the Los Alamos or University of California-San  
 Diego libraries that are more significantly related to the amino  
 acid nucleic acid sequence of v-rel than to the randomized  
 seq of v-rel. Comparison of Rev-A, Rev-T, and c-rel indicates  
 that c-rel sequences may have been transduced from the c-rel  
 (turkey) locus by a novel mechanism. There are sequences in Rev-A  
 and c-rel that are similar to splicing signals, indicating that the  
 5' c-rel junction of Rev-T may have been formed by cellular RNA  
 spl machinery. Eight presumed introns have presumably been  
 spl out of c-rel to generate v-rel. There are also short  
 imp regions of homology between sequences at the boundaries of  
 v-rel and c-rel sequences in Rev-A and c-rel (turkey), indicating that  
 c-rel sequences have been transduced by **homologous**  
**recombination**. There are many differences between the amino  
 acid sequences of the predicted translational products of v-rel and  
 c-rel which may account for their difference in transformation  
 pot. These sequence differences between v-rel and c-rel  
 incl missense transitions, four missense transversions, and  
 thr deletions where Rev-T has a small in-frame deletion of sequences  
 rel to c-rel. Most of the coding sequence differences between  
 c-rel and v-rel are nonconservative amino acid changes.

L9 ANSWERS OF 36 MEDLINE  
 AN 842 MEDLINE  
 DN 842  
 TI Rec Transfection of transfected DNAs in vertebrate cells in culture.  
 AU Banerjee P K; Watanabe S; Temin H M  
 NC CA- (NCI)  
 CA- (NCI)  
 SO PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES  
 OF AMERICA (1984 Jun) 81 (11) 3476-80.  
 Jou Title: PV3. ISSN: 0027-8424.  
 CY Uni Titles  
 DT Journal Article; (JOURNAL ARTICLE)  
 LA Eng  
 FS Pri Journals; Cancer Journals  
 EM 198

AB We the frequency of homologous and illegitimate  
 rec ons between transfected noninfectious **retroviral**  
 DNA les in chicken embryo fibroblasts. The frequency of  
 rec on was determined by the formation of infectious virus  
 and with the extent of homology between the DNA molecules at  
 the recombination, but only when there were regions of  
 hom surrounding this region. While **homologous**  
 recombination led to the formation of wild-type virus,  
 illegitimate recombination resulted in formation of infectious virus  
 with mutations at the site of recombination. Apparent  
**homologous recombination** was also observed between  
 tra and **chromosomal** DNAs in D17 dog cells.

L9 ANS OF 36 MEDLINE  
 AN 840 MEDLINE

DN 840  
 TI Int osomal recombination of the cellular oncogene c-myc with  
 the globulin heavy chain locus in murine plasmacytomas is a  
 rec exchange.

AU Cor rondakis S; Adams J M  
 NC CA (ICI)  
 SO EMB AL, (1983) 2 (5) 697-703.  
 Jou le: EMB. ISSN: 0261-4189.  
 CY ENG nited Kingdom  
 DT Jou rticle; (JOURNAL ARTICLE)

LA Eng  
 FS Pri ournals  
 EM 198

AB The chromosome translocations found in most murine  
 pla mas involve the cellular gene (c-myc) homologous to the  
 onc (c-myc) of avian **retrovirus** MC29, Translocation  
 lin c-myc gene of **chromosome** 15 to the  
 imm ulin heavy (H) chain locus of **chromosome** 12,  
 oft in the switch recombination (S) region 5' to the alpha  
 con region (C alpha) gene. We have investigated c-myc  
 rea ents in 21 BALB/c plasmacytomas and three B lymphomas by  
 Sou ot analysis. We show that the t(15;12) is a reciprocal  
**chr** exchange since most tumours contain not only a  
 c-m linked to the S alpha C alpha region but also a separate  
 str with S mu or S alpha linked to the c-myc 5'-flanking  
 reg nalysis of the two rearrangement products cloned from  
 pla na J558 suggests that one type of H locus target for  
 tra on is an S alpha region recombined with S mu; two other  
 tar pear to be other switched heavy chain genes and an  
 unr ed C alpha gene. Nearly all the **chromosome** 15  
 bre s fall within a 1.1-kb region spanning a 5' c-myc exon;  
 hen sion of the transcriptional unit by translocation can  
 acc of the altered c-myc transcription in plasmacytomas. The  
 c-m ppoint region lacks substantial homology with S mu or S  
 alpa ing against **homologous recombination**  
 as nslocation mechanism.

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